



King's Research Portal

DOI:

[10.7554/eLife.59272](https://doi.org/10.7554/eLife.59272)

Document Version

Publisher's PDF, also known as Version of record

[Link to publication record in King's Research Portal](#)

Citation for published version (APA):

Jager, P., Moore, G., Calpin, P., Durmishi, X., Salgarella, I., Menage, L., Kita, Y., Wang, Y., Kim, D. W., Blackshaw, S., Schultz, S. R., Brickley, S., Shimogori, T., & Delogu, A. (2021). Dual midbrain and forebrain origins of thalamic inhibitory interneurons. *eLife*, 10, 1-29. [e59272]. <https://doi.org/10.7554/eLife.59272>

Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

Take down policy

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Dual midbrain and forebrain origins of thalamic inhibitory interneurons

Polona Jager¹, Gerald Moore^{2,3}, Padraic Calpin⁴, Xhuljana Durmishi¹, Irene Salgarella¹, Lucy Menage¹, Yoshiaki Kita⁵, Yan Wang⁵, Dong Won Kim⁶, Seth Blackshaw⁶, Simon R Schultz², Stephen Brickley³, Tomomi Shimogori⁵, Alessio Delogu^{1*}

¹Department of Basic and Clinical Neuroscience, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London, United Kingdom; ²Department of Bioengineering, Imperial College London, London, United Kingdom; ³Department of Life Sciences and Centre for Neurotechnology, Imperial College London, London, United Kingdom; ⁴Department of Physics and Astronomy, University College London, London, United Kingdom; ⁵RIKEN, Center for Brain Science (CBS), Saitama, Japan; ⁶The Solomon H. Snyder Department of Neuroscience, School of Medicine, Johns Hopkins University, Baltimore, United States

Abstract The ubiquitous presence of inhibitory interneurons in the thalamus of primates contrasts with the sparsity of interneurons reported in mice. Here, we identify a larger than expected complexity and distribution of interneurons across the mouse thalamus, where all thalamic interneurons can be traced back to two developmental programmes: one specified in the midbrain and the other in the forebrain. Interneurons migrate to functionally distinct thalamocortical nuclei depending on their origin: the abundant, midbrain-derived class populates the first and higher order sensory thalamus while the rarer, forebrain-generated class is restricted to some higher order associative regions. We also observe that markers for the midbrain-born class are abundantly expressed throughout the thalamus of the New World monkey marmoset. These data therefore reveal that, despite the broad variability in interneuron density across mammalian species, the blueprint of the ontogenetic organisation of thalamic interneurons of larger-brained mammals exists and can be studied in mice.

*For correspondence:
alessio.delogu@kcl.ac.uk

Competing interests: The authors declare that no competing interests exist.

Funding: See page 23

Received: 24 May 2020

Accepted: 31 January 2021

Published: 01 February 2021

Reviewing editor: Sonia Garel, Ecole Normale Supérieure, France

© Copyright Jager et al. This article is distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use and redistribution provided that the original author and source are credited.

Introduction

The thalamus is a forebrain structure that develops from the diencephalic prosomere 2 (p2) (Puelles and Rubenstein, 2003; Shi et al., 2017; Wong et al., 2018) and is primarily composed of cortically projecting excitatory thalamocortical (TC) neurons, divided into more than 30 individual nuclei in mammals (Clascá et al., 2012; Hunnicutt et al., 2014; Jones, 2007). The function of the thalamus has been historically described as relay of sensory information to the cortex (Cheong et al., 2013; Hubel and Wiesel, 1962; Piscopo et al., 2013; Shatz, 1996; Sherman and Guillery, 2002; Van der Loos and Woolsey, 1973; Zeater et al., 2015). Taking into account the diversity of input and output features of TC circuits (Clascá et al., 2012; Guillery, 1995; Herkenham, 1980; Rubio-Garrido et al., 2009; Sherman, 2016), more recent work has shown that the thalamus is also critically involved in cognitive processes allowing for behavioural flexibility (Bolkan et al., 2017; Groh et al., 2014; Guo et al., 2017; Ling et al., 2015; Rikhye et al., 2018a; Rikhye et al., 2018b; Saalman and Kastner, 2011; Schmitt et al., 2017; Sherman, 2016).

In contrast to cortical networks, excitatory neurons in the thalamus do not connect with each other (Bickford et al., 2008; Hirsch et al., 2015; Jones, 2007; Rikhye et al., 2018b). Instead, local connections and computations within TC circuits are dominated by the resident inhibitory, GABA-

releasing neurons (interneurons) (Hirsch et al., 2015; Montero, 1987; Pasik and Pasik, 1976; Sherman, 2004).

Interneuron numbers and distribution vary widely across species, suggesting that they are critically involved in the evolution of TC structure and function (Arcelli et al., 1997; Letinic and Rakic, 2001; Rikhye et al., 2018b). In particular, comparative studies across all amniotes (reptiles, birds, and mammals) have described a correlation between the proportion of interneurons and the size and connectivity of the excitatory thalamus (Arcelli et al., 1997; Butler, 2008).

For example, in the reptilian thalamus, which is mostly devoid of descending projections from the cortex, interneurons have only been identified in the retinorecipient regions (Butler, 2008; Kenigfest et al., 1995; Kenigfest et al., 1998; Pritz and Stritzel, 1994; Rio et al., 1992). In birds, however, where reciprocal connections between the thalamus and the cortex are more abundant, thalamic interneurons are distributed more widely (Butler, 2008; Granda and Crossland, 1989; Veenman and Reiner, 1994).

Similarly among mammals, interneurons are largely restricted to the visual thalamus in smaller-brained marsupials, bats and mice, where they represent only 6% of the total neuronal population (Butler, 2008; Evangelio et al., 2018; Seabrook et al., 2013b). In primates, on the other hand, where higher order (HO) nuclei driven by cortical inputs are expanded relative to sensory relay (first order, FO) regions (Armstrong, 1979; Baldwin et al., 2017; Butler, 2008; Halley and Krubitzer, 2019; Stephan et al., 1981), interneurons are present across the entire thalamus and their proportion increases to around 30% (Arcelli et al., 1997; Braak and Bachmann, 1985).

To what extent these differences are the result of species-specific ontogenesis of thalamic interneurons remains poorly understood. We have previously shown that in the mouse, interneurons in the FO visual thalamus, the dorsal lateral geniculate nucleus (LGd), originate in the midbrain from an *En1⁺Gata2⁺Otx2⁺Sox14⁺* lineage (Jager et al., 2016). On the other hand, earlier work in humans has suggested the DLX1/2-expressing ganglionic eminences (GE) in the telencephalon as the source of interneurons for some of the larger HO thalamic nuclei – the mediodorsal (MD) nucleus and the pulvinar (Letinić and Kostović, 1997; Letinic and Rakic, 2001; Rakić and Sidman, 1969). At the same time, the latter studies were not able to detect any such migration from the GE in the mouse and macaque brain (Letinic and Rakic, 2001). While these findings therefore point to innovation in developmental origins, we currently lack an understanding of the shared ontogeny of mammalian thalamic interneurons.

Here we hypothesised that a blueprint of the complex organisation of thalamic interneurons observed in large-brained mammals is present even in the simpler thalamus of the mouse. This prediction is supported by findings from the cortex demonstrating that its inhibitory interneuron classes, generated in the subpallium and defined through expression of regulatory programmes (i.e. transcription factors), are common to the amniote lineages (Arendt et al., 2019; Metin et al., 2007; Tasic et al., 2018; Tosches et al., 2018). Moreover, a conserved subpallial origin was demonstrated for cortical interneurons in the cyclostome hagfish, and therefore appears to be an ancestral feature of the vertebrate brain (Sugahara et al., 2017; Sugahara et al., 2016).

Using genetic fate mapping, two-photon whole brain tomography, and spatial modelling, we investigated the ontogeny and distribution of thalamic GABAergic interneurons comprehensively across the mouse TC nuclei. We then used fluorescent in situ marker detection to compare the distribution of genetically defined interneuron classes in the New World marmoset monkey brain. These experiments identify in the mouse a wider distribution of GABAergic interneurons than previously reported (Arcelli et al., 1997; Evangelio et al., 2018; Seabrook et al., 2013b), encompassing both FO sensory relay and HO thalamic nuclei, including associative HO nuclei that are most enlarged in primates. We then show that while the largest proportion of thalamic interneurons in the mouse is generated in the *En1⁺Sox14⁺* embryonic midbrain, there is an additional class that derives from the *Nkx2.1⁺Lhx6⁺Dlx5⁺Foxd1⁺* inhibitory progenitor domains in the forebrain. Intriguingly, we also find that in the mouse interneurons are organised in a spatial pattern according to their ontogeny, such that midbrain-born interneurons are largely found in the sensory relays, while the forebrain-generated interneurons reside in the HO thalamus, including MD, laterodorsal (LD), and lateral posterior (LP; aka pulvinar) nuclei. Genoarchitectonic evidence supports a conserved basic organisation of thalamic interneurons in the non-human primate marmoset thalamus, where putative midbrain-generated interneurons are abundant in FO and HO nuclei and complemented by a distinct and more restricted interneuron class enriched in selected HO associative TC nuclei.

Results

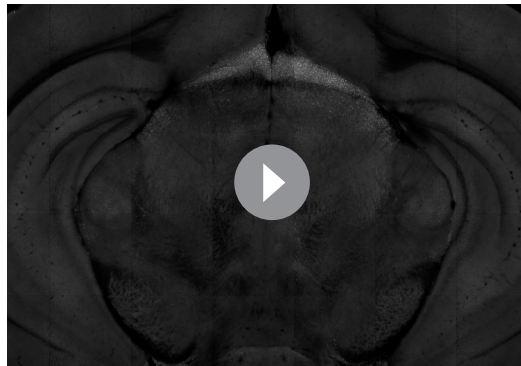
Sox14-expressing interneurons are widely distributed across the FO and HO mouse thalamus

In the mouse thalamus, GABAergic interneurons are most abundant in the LGd (Arcelli et al., 1997; Evangelio et al., 2018). We had previously demonstrated that all LGd interneurons are defined by expression of the transcription factor gene Sox14 and presented the Sox14^{GFP/+} knockin mouse line (Table 1; Crone et al., 2008) as a useful tool to study these cells (Jager et al., 2016). Both the Allen Brain Atlas (2015 Allen Institute for Brain Science. Allen Cell Types Database. Available from: cell-types.brain-map.org) and DropViz resources (available from: dropviz.org; Saunders et al., 2018) identify a Sox14⁺ transcriptional cluster corresponding to mouse LGd interneurons, confirming our previous findings. Sox14 is expressed upon cell-cycle exit within inhibitory lineages in the diencephalon, midbrain, hindbrain, and spinal cord, but not in the telencephalon (Achim et al., 2013; Delogu et al., 2012; Guo and Li, 2019; Prekop et al., 2018).

To investigate the spatial distribution of Sox14 neurons comprehensively across all thalamic (TC) regions in the mouse, we took advantage of the endogenous and bright fluorescence of GFP in postnatal day (P) 21 Sox14^{GFP/+} mice to perform high resolution (0.54 μm voxel) whole brain imaging by two-photon laser scanning tomography; for an indicative low-resolution scan through a series of z-projections, see also: Video 1. Optical sections were acquired in the coronal plane 10 μm apart and registered with the Allen Institute Common Coordinate Framework (CCF3; Figure 1A) using a custom Python pipeline alongside the Elastix registration library (Klein et al., 2010) to delineate anatomical subdivisions according to the Allen Brain Institute taxonomy (Figure 1A and Video 2). To perform automated detection of cells, a deep learning strategy was implemented to train a U-Net segmentation model (Ronneberger and Brox, 2015) on a data set of 12,264 images (512 × 512 pixels at 0.54 μm voxel), obtained by supplemental augmentation of 219 manually annotated samples (Figure 1B). The accuracy of our automated counting strategy was validated by comparing the total count of GFP⁺ cells in the LGd of P21 Sox14^{GFP/+} mice (Figure 1C,D; 1234 ± 82; mean ± SD) to a recent stereological study of LGd GABA interneurons in the wild-type adult C57Bl/6 mouse (1255 ± 195; mean ± SD) (Evangelio et al., 2018), which confirmed the validity of our protocol.

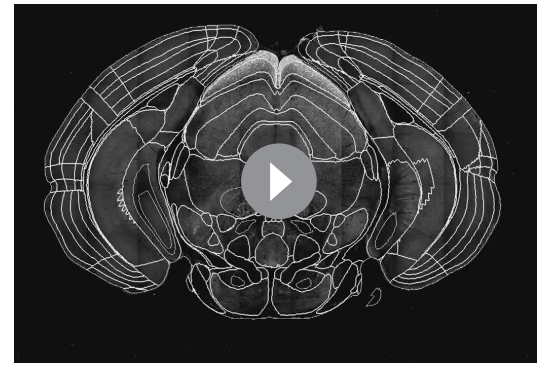
Table 1. Animal models used in the study.

Species	Designation	Source or reference	Identifiers	Additional information
Mus musculus	Sox14 ^{tm1Tmj} (Sox14 ^{GFP})	Crone et al., 2008	MGI ID: 3836003	Maintained in the C57BL/6J (Charles River Laboratories) background
Mus musculus	En1 ^{Cre}	Kimmel et al., 2000; The Jackson Laboratory	Stock No: 007916 MGI ID: 2446434	C57BL/6J background
Mus musculus	Dlx5/6 ^{Cre}	Monory et al., 2006; The Jackson Laboratory	Stock No: 008199; MGI ID:3758328	C57BL/6J background
Mus musculus	Pvalb ^{Cre}	Hippenmeyer et al., 2005; The Jackson Laboratory	Stock No: 017320; MGI ID:3590684	C57BL/6J background
Mus musculus	Nkx2.1 ^{Cre}	Xu et al., 2008	Stock No: 008661 MGI: J:131144	C57BL/6J background
Mus musculus	Foxd1 ^{EGFPcre} (Foxd1 ^{Cre})	Humphreys et al., 2008	Stock No: 012463 MGI:4359653	C57BL/6J background
Mus musculus	Lhx6 ^{Cre}	Fogarty et al., 2007	Stock No: 026555 MGI:4355717	C57BL/6J background
Mus musculus	RCE:loxP (Rosa26 ^{Isl-GFP})	Sousa et al., 2009; The Jackson Laboratory	MMRRC Stock No: 32037-JAX MGI:4412373	C57BL/6J background
Mus musculus	Gt(ROSA)26Sortm5(CAG-Sun1/sfGFP)Nat (Rosa26 ^{Isl-nuclearGFP})	Mo et al., 2015; The Jackson Laboratory	Stock No: 021039; MGI ID: 5443817	C57BL/6J background
Callithrix jacchus	WT	Colony at RIKEN RRD	-	-



Video 1. A video animation of 28 z-stacks (each 100 μm) of projected coronal slices, downsized to 1 μm voxel size in XY to reduce file size. Each slice is a maximum intensity projection across 10 serial two-photon optical slices.

<https://elifesciences.org/articles/59272#video1>



Video 2. A video animation of the anatomical delineations from the Allen Institute Common Coordinate Framework (CCF3) projected onto the imaging data from a P21 *Sox14^{GFP/+}* brain. Isotropic 10 μm voxel size.

<https://elifesciences.org/articles/59272#video2>

Automated counting identified a total of 6588 ± 811 (mean \pm SEM) *GFP⁺* cells across TC nuclei of both hemispheres spanning the rostrocaudal and mediolateral extensions of the thalamus ($n = 3$ *Sox14^{GFP/+}* at P21). Their distribution was not stochastic but skewed instead towards sensory TC nuclei and within sensory modality by a prevalence in FO nuclei. The *GFP⁺* cells are most abundant in the visual FO LGd (1234 ± 82) and HO LP (aka pulvinar; 411 ± 110), followed by sensory-motor ventrobasal (VB) complex [ventral posteromedial (VPM; 250 ± 49), ventral posterolateral (VPL; 105 ± 40), ventral anterior lateral (VAL; 4 ± 3) and ventral medial nucleus (VM; 4 ± 2)] and HO posterior nucleus (PO; 99 ± 12), in turn followed by the auditory FO ventral medial geniculate (MGv; 53 ± 12) and HO dorsal medial geniculate nucleus (MGd; 20 ± 13). Sparse *GFP⁺* cells were also detected in limbic TC nuclei: LD (20 ± 7), parafascicular (PF) (25 ± 5), and MD (13 ± 6) nuclei. Notably, *GFP⁺* neurons are typically found in the caudal-most part of the LD, nearer the LP border and nearly all *GFP⁺* neurons were in the caudal half of the MD (see also: **Video 1**). Counts per TC nucleus are reported for a single hemisphere.

Sox14 expression distinguishes between two spatially clustered interneuron classes

To validate the inhibitory nature of the *GFP⁺* cells in TC nuclei of the *Sox14^{GFP/+}* mouse, we combined immunodetection of *GFP⁺* with in situ RNA hybridisation (ISH) against the *Gad1* mRNA in *Sox14^{GFP/+}* mice sampling the rostrocaudal extent of the thalamus at approximately 200 μm intervals (**Figure 2A–C**). In our experience, the simultaneous detection of protein and mRNA is more reliable in younger tissue, hence experiments were done at P14, by which time point mouse TC nuclei are considered to display adult-like circuit composition (*Bickford et al., 2010*; *Golding et al., 2014*; *Seabrook et al., 2013a*; *Seabrook et al., 2013b*; *Thompson et al., 2016*).

Recapitulating the distribution observed at P21, *GFP⁺* cells were detected in the LGd, LP, VP, and PO and in very small numbers in the MG (**Figure 2A,C,D**). In these nuclei all *GFP⁺* cells had a GABAergic profile and co-expressed *Gad1* (100%, $n = 3$ brains). In the LGd, VP, and MGv (i.e. FO sensory relay nuclei) they also represented virtually all GABAergic cells ($\geq 98\%$, pie charts in **Figure 2D**).

Unexpectedly, however, $22.1 \pm 4.0\%$ of the total GABAergic population in TC regions did not express *GFP* (**Figure 2B,C**; **Figure 5Bii**), and these *GFP⁻Gad1⁺* cells appeared spatially largely non-overlapping with the *Sox14⁺* interneuron class and enriched at more rostral TC territory (**Figure 2C,D**). *Gad1⁺* cells are seen populating these rostral TC regions during the first five postnatal days (**Figure 2—figure supplement 1**). In particular, we observed that the distribution of *GFP⁻Gad1⁺* cells is skewed towards the limbic HO MD ($29.6 \pm 4.5\%$) and LD ($19.4 \pm 3.1\%$) and the associative HO LP ($13.2 \pm 1.2\%$), and in smaller numbers towards the PO. Sparse *GFP⁻Gad1⁺* cells were also found in thalamic regions where nuclear boundaries cannot be defined precisely at this age and that contain

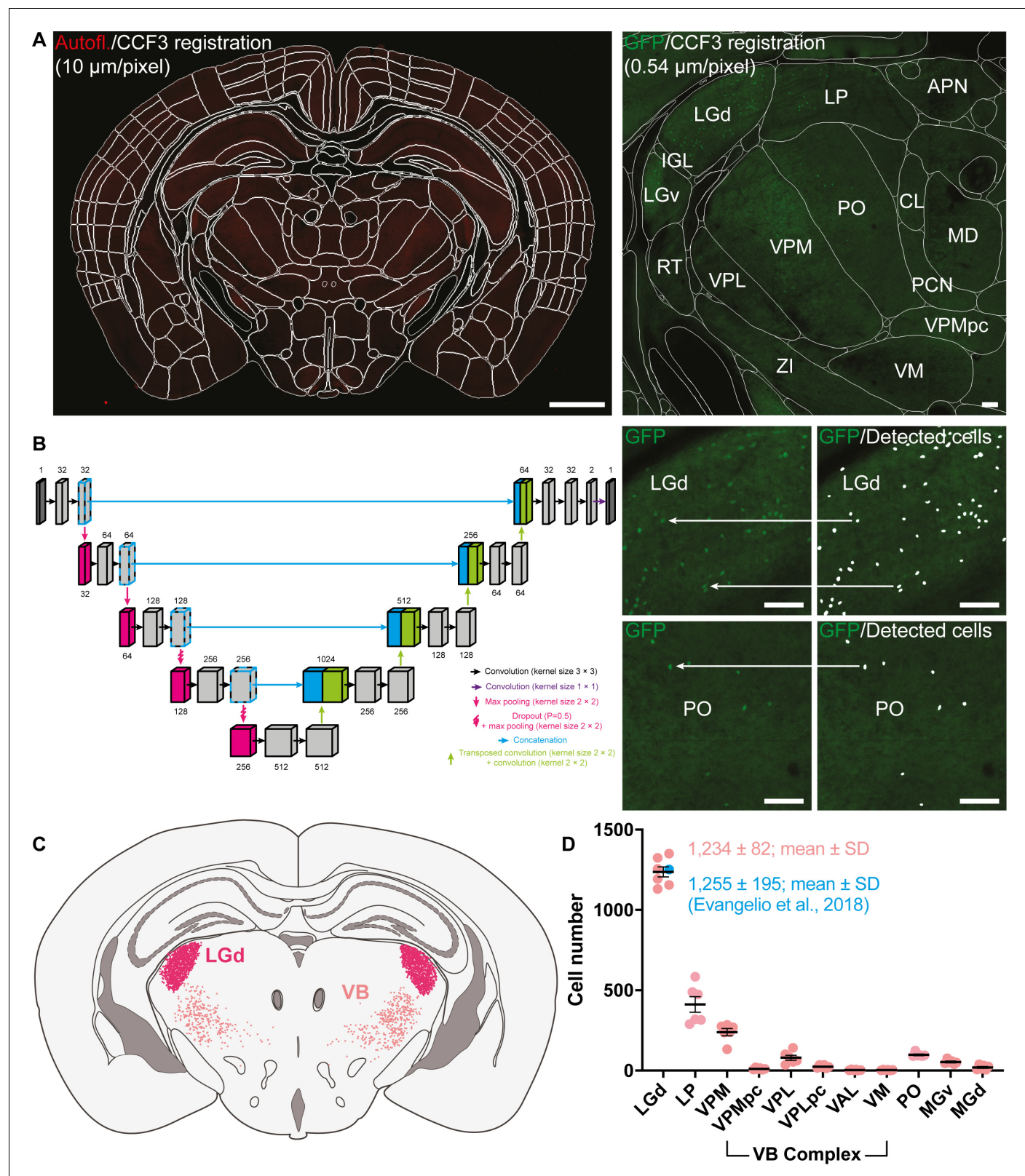


Figure 1. Automated total counts of GFP⁺ cells in the thalamus of the Sox14^{GFP/+} mouse. (A) Autofluorescence (Autofl.) from serial two-photon imaging of Sox14^{GFP/+} mice (n = 3) at 0.54 \times 0.54 \times 10 μm voxel resolution was registered to the Allen Institute CCF3 atlas using Elastix (left; scale bar 1 mm). This permits delineation and identification of all anatomical structures according to the Allen Institute hierarchical taxonomy (right; scale bar 100 μm). (B) Automated cell detection was done using a U-Net trained on 219 manually segmented images (512 \times 512 pixels) augmented to a total sample size

Figure 1 continued on next page

Figure 1 continued

of 12,264, split 75% for training and 25% validation. Images containing GFP fluorescence were passed into the trained U-Net (left) for cell prediction based on learned features during training (right; scale bar 100 μ m). Oversampling in the z-axis was corrected for by grouping and averaging detected cell positions which colocalised within a defined cell radius. (C) Example illustration of automatically detected cells in the LGd and VB complex projected onto a representative coronal section of the thalamus. (D) Quantification of GFP⁺ cells in the LGd at 1234 ± 82 (mean \pm SD) validated against stereological study by [Evangelio et al., 2018](#) of 1255 ± 195 (mean \pm SD) interneurons in the LGd. Other counts are shown for LP, VB complex [VPM, parvicellular part of the ventral posteromedial nucleus (VPMpc), VPL, parvicellular part of the ventral posterolateral nucleus (VPLpc), VAL, VM], MGv, MGd, and PO.

VAL, VM, centrolateral (CL), intermediodorsal (IMD), PF, reuniens (RE), rhomboid (RH), subparafascicular (SPF), subparafascicular area (SPA), central medial (CM), and anteromedial (AM) nuclei. Cell counts in these regions were grouped together under 'other TC' ([Figure 2D](#)).

To quantitatively demonstrate spatial clustering of these two putative thalamic interneuron classes ($Gad1^+Sox14^+$ and $Gad1^+Sox14^-$), we calculated the nearest neighbour distances (NND) from 3D reconstructions of their respective distributions in the $Sox14^{GFP/+}$ thalamus ([Figure 2C](#) and [Figure 3A](#)). Indeed, the cumulative distribution of NNDs was significantly shifted to smaller distances within each of the classes than between them ($p < 1.4 \times 10^{-30}$, two-sample Kolmogorov–Smirnov test, $n = 3$ brains; [Figure 3A](#)).

To characterise spatial organisation of thalamic GABAergic interneurons in an unbiased way, we then applied machine learning (k-means clustering) to these same 3D reconstructions of the $Sox14^{GFP/+}$ thalami ([Figures 2C](#) and [3B,C](#)). The data best fit two spatial clusters, as assessed from the silhouette score ([Figure 3Bii,C](#); see also Materials and methods). Consistent with the NND analysis, one cluster corresponded to the $Sox14^+$ cells (contains $94.9 \pm 1.4\%$ of all $Sox14^+$ cells), and the other to the $Sox14^-$ interneurons (contains $81.0 \pm 0.3\%$ of all $Sox14^-$ cells; [Figure 3B,D](#)). The two thalamic molecular GABAergic groups therefore occupy their own respective spatial clusters, with the $Sox14^+$ cells located more rostrally and medially compared to the $Sox14^-$ interneurons.

To independently confirm our findings and control for potential effects of looking at a juvenile age (P14), we also mapped anatomical distribution of all $Gad1^+$ and $Chrna6^+$ cells across the adult mouse TC nuclei at P56, using the Allen Mouse Brain Atlas (2004 Allen Institute for Brain Science. Allen Mouse Brain Atlas. Available from: mouse.brain-map.org; [Lein et al., 2007](#)) ISH data ([Figure 3—figure supplement 1A](#)). *Chrna6* has been identified as another marker specific for interneurons, at least in the LGd ([Golding et al., 2014](#); DropViz; Allen Cell Types Database). The resulting 3D reconstructions, k-means spatial clustering ([Figure 3—figure supplement 1A](#)), and distribution plot ([Figure 3—figure supplement 1B](#)) were consistent with our observations from the P14 $Sox14^{GFP/+}$ thalamus.

The mouse thalamus therefore exhibits wider interneuron diversity than has been previously reported, with at least two molecularly and spatially distinct classes. The largest interneuron class is defined by $Sox14^+$ and is enriched in the caudal part of the thalamus which contains principal sensory relays and their associated HO nuclei. Conversely, the smaller $Sox14^-$ GABAergic population is enriched in the rostral part of the thalamus, in HO regions that associate with more cognitive functions, such as the MD and LD ([Halassa and Kastner, 2017](#); [Rikhie et al., 2018b](#)).

The $Sox14^+$ interneuron class is abundant and widespread in the marmoset thalamus

Given the sparseness of interneurons in the mouse thalamus, there exists the possibility that the $Sox14^+$ interneuron class may represent a unique feature of smaller-brained species, or that it may be a conserved, but numerically negligible type of interneuron complemented by novel and more abundant types in species with larger brains. To detect the presence and assess the relative abundance of the $Sox14^+$ interneuron class in the thalamus of species with a high density of interneurons, we sampled the distribution of $SOX14^+GAD1^+$ cells in selected TC nuclei of the neonatal non-human primate marmoset.

Fluorescent ISH for *SOX14* and *GAD1* mRNAs revealed the widespread presence of $SOX14^+GAD1^+$ bona fide interneurons across all major TC nuclei ([Figure 4A–C](#)). Reminiscent of the expression pattern observed in the mouse, $SOX14^+$ cells were not present in prethalamic structures [RT,

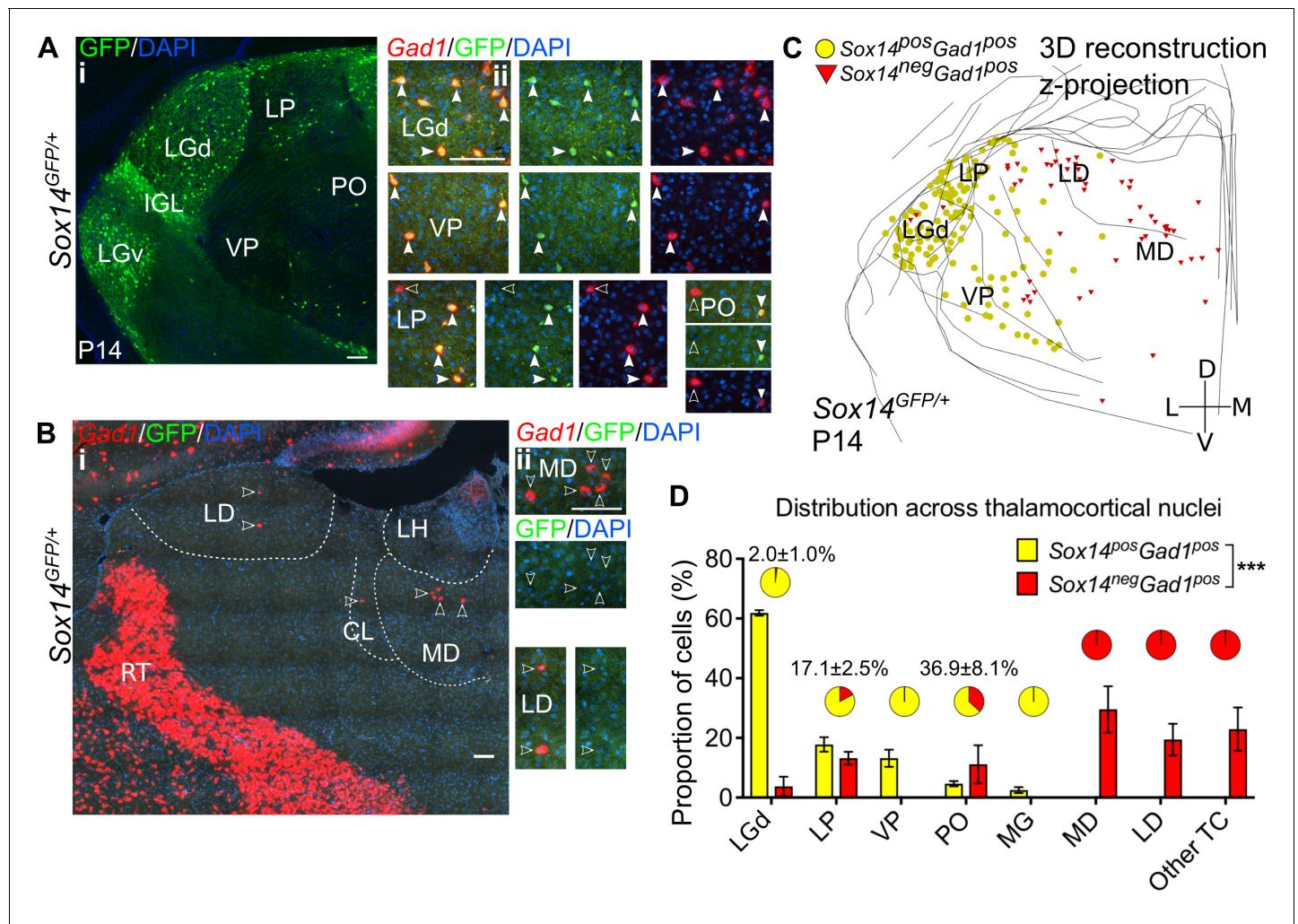


Figure 2. Diversity and distribution of GABAergic cells in the mouse thalamocortical nuclei. (A) (i) Representative coronal section of P14 *Sox14*^{GFP/+} thalamus with *Sox14*⁺ cells in the LGd, VP, LP, and PO. (ii) *Sox14*⁺ cells in TC regions co-express *Gad1*, but not all *Gad1*⁺ cells co-express *Sox14* in the LP and PO. Filled arrows mark *Sox14*⁺*Gad1*⁺ and empty arrows *Sox14*⁻*Gad1*⁺ cells. Scale bars, 100 μ m. (B) (i) Representative rostral coronal section of P14 *Sox14*^{GFP/+} thalamus with *Gad1*⁺ cells in the MD, CL, and LD, which contain no *Sox14*⁺ cells. (ii) *Gad1*⁺ cells in these nuclei do not co-express *Sox14*. Scale bars, 100 μ m. (C) 3D reconstruction of a representative P14 *Sox14*^{GFP/+} thalamus from tracing every tenth 20- μ m-thick coronal section, displayed as a z-projection and showing distribution of *Sox14*⁺*Gad1*⁺ (yellow) and *Sox14*⁻*Gad1*⁺ cells (red). One dot represents one neuron. (D) Distribution of *Sox14*⁺*Gad1*⁺ and *Sox14*⁻*Gad1*⁺ cells across TC nuclei in the *Sox14*^{GFP/+} brains at P14, plotted as proportion of all the cells within each interneuron group (mean \pm SEM; $n = 3$ brains). The category 'other TC' refers to regions where nuclear boundaries cannot be defined precisely and that contain VAL, VM, CL, IMD, PF, RE, RH, SPF, SPA, CM, and AM. *Sox14*⁺*Gad1*⁺ and *Sox14*⁻*Gad1*⁺ populations have distinct distributions ($p < 10^{-3}$ chi-squared test). Pie charts show the proportion (mean \pm SEM) of the two interneuron classes within each nucleus. The online version of this article includes the following figure supplement(s) for figure 2:

Figure supplement 1. Perinatal distribution of *Gad1*⁺ cells in the anterior thalamus.

zona incerta (ZI)], but detectable in the pregeniculate/subgeniculate (SubG), the primate homologue of the mouse IGL (**Figure 4B**).

Qualitative analysis shows largely overlapping distribution of the fluorescent probes for SOX14 and GAD1 in all TC nuclei at caudal (**Figure 4Ai-vi**) and intermediate levels (**Figure 4Bi-iii**), but some areas of differential expression at rostral level, where GAD1 expression is not accompanied by SOX14 expression in medial and dorsal regions of the thalamus (**Figure 4C**) that contain the limbic HO TC nuclei LD (**Figure 4Ci**) and MD (**Figure 4Cii**). We then proceeded to quantify the number of SOX14⁺GAD1⁺ and SOX14⁻GAD1⁺ cells in three brains of the new-born marmoset by randomly selecting three regions of 263 μ m by 263 μ m within the following nuclei: FO visual LGd, HO visual

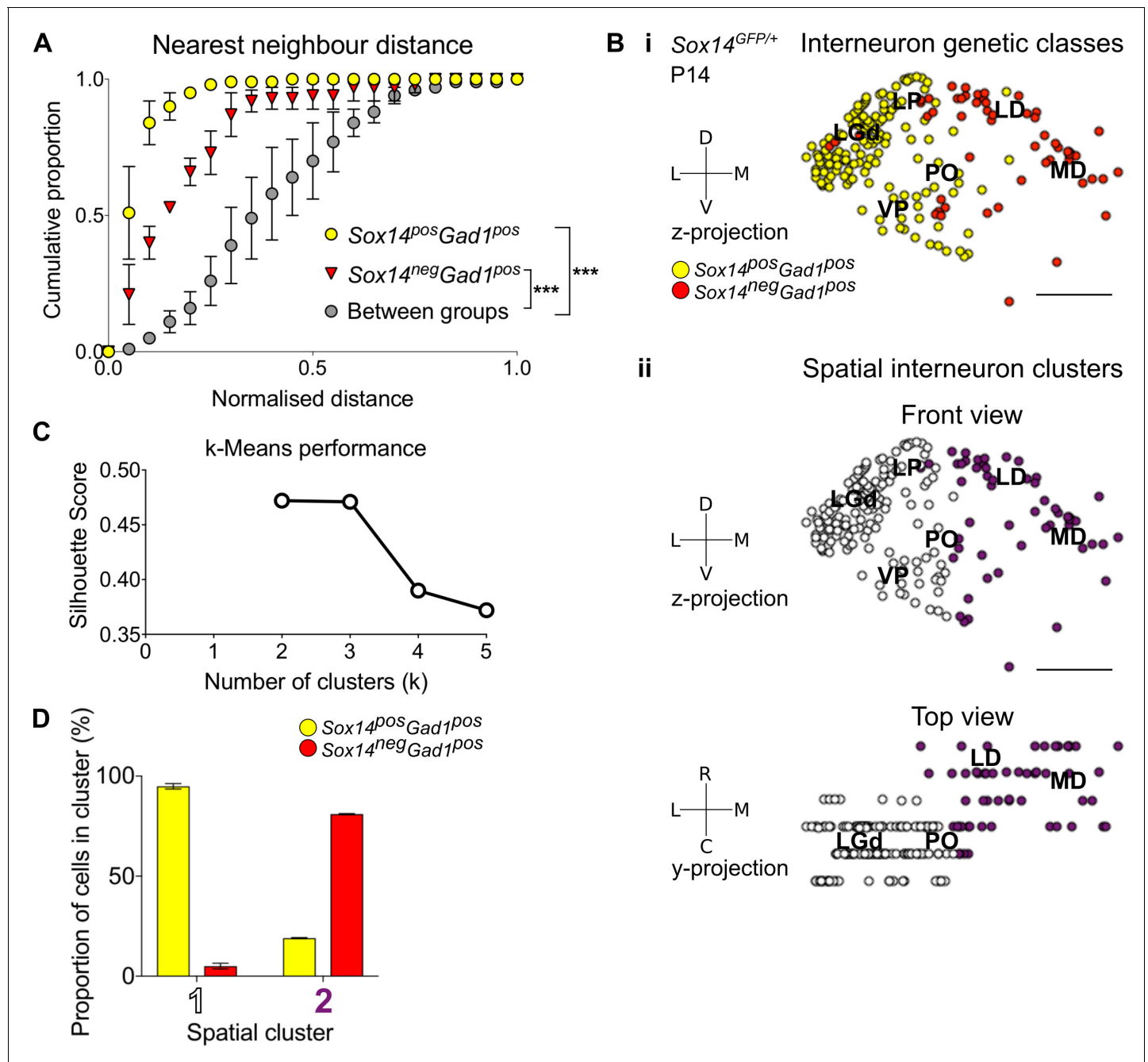


Figure 3. Spatial organisation of thalamic GABAergic cells. (A) Normalised nearest neighbour distance (NND) for $Sox14^{+}Gad1^{+}$ and $Sox14^{-}Gad1^{+}$ populations and between the two groups from P14 $Sox14^{GFP/+}$ data (Figure 2), plotted as cumulative proportion of all cells within a given set. The NND distribution is significantly shifted to larger distances between groups than within each of the groups ($p < 1.4 \times 10^{-30}$, two-sample Kolmogorov–Smirnov test, $n = 3$ brains). (B) Representative z-projections of interneuron distribution amongst TC nuclei, from P14 $Sox14^{GFP/+}$ data (Figure 2). One dot represents one neuron and they are colour coded by (i) their genetic identity or (ii) spatial cluster. For the spatial clusters a y-projection is also shown. Scale bars, 500 μm . (C) Performance of unsupervised k-means algorithm in identifying thalamic interneuron spatial clusters from the P14 $Sox14^{GFP/+}$ data ($n = 3$ brains, see also Figure 2) as measured by the silhouette score, which varies with number of clusters (k). We choose $k = 2$ as this point has the highest score. (D) Proportion of $Sox14^{+}$ and $Sox14^{-}$ GABAergic cells in each spatial cluster, averaged over three brains (mean \pm SEM). The online version of this article includes the following figure supplement(s) for figure 3:

Figure supplement 1. Spatial organisation of $Gad1^{+}$ and $Chrna6^{+}$ cells in the adult mouse thalamus.

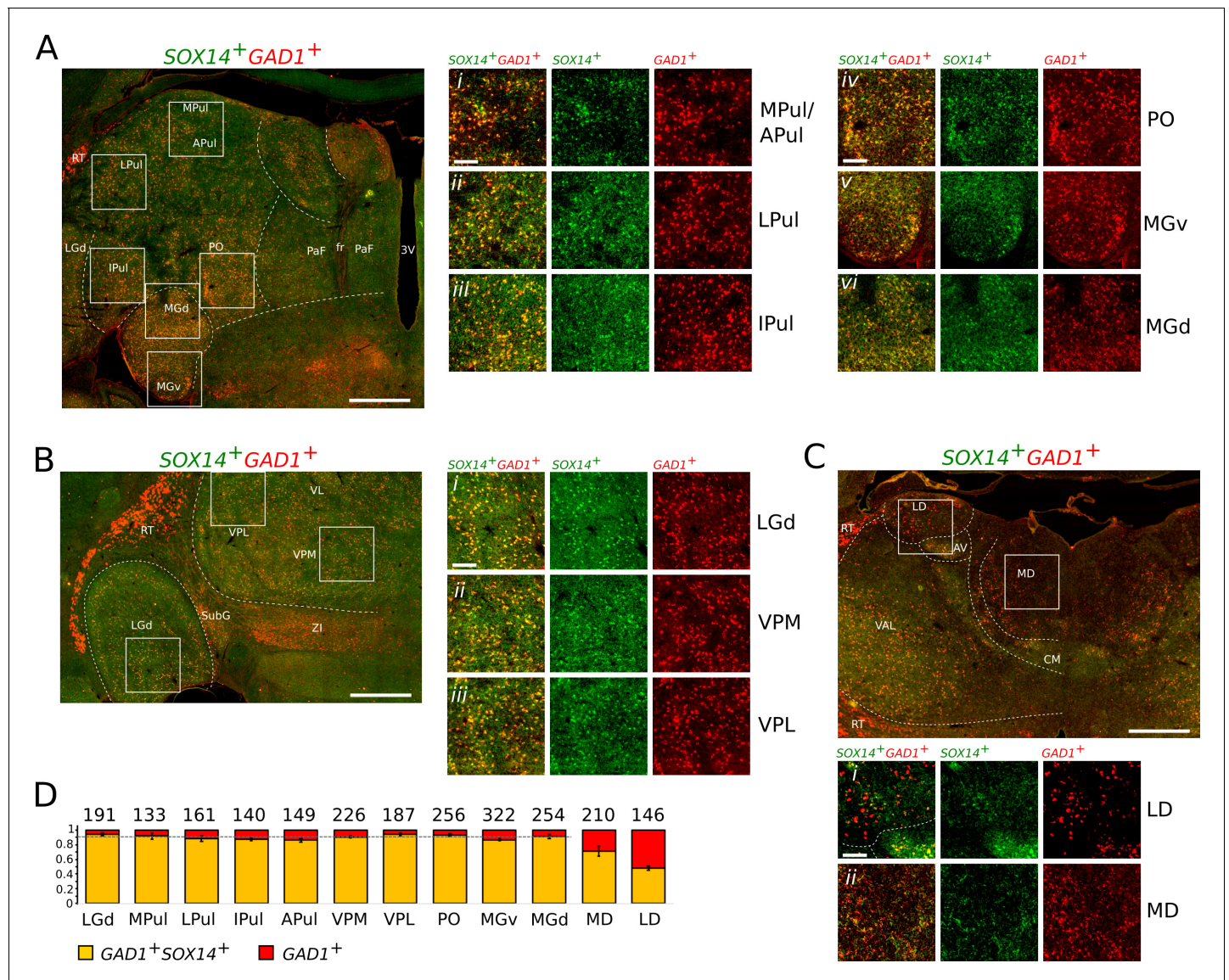


Figure 4. SOX14⁺GAD1⁺ interneurons dominate TC regions of the non-human primate marmoset. Representative coronal sections of the thalamus of a new-born marmoset illustrating the distribution of cells expressing the SOX14 (green) and GAD1 (red) mRNAs. (A) Caudal plane containing subdivisions of the pulvinar complex, the PO, and the auditory MG. Also visible are parafascicular (PaF) nuclei. Fr, fasciculus retroflexus; 3V, third ventricle. (Ai–iii) Magnifications of indicative areas of the medial (MPul) and anterior pulvinar (APul), the lateral pulvinar (LPul), and inferior pulvinar (IPul). (Aiv). Magnification of a region of the PO. (Av and vi). Magnifications of representative regions of the ventral (MGv) and dorsal (MGd) subdivisions of the auditory thalamus. (B) Middle plane section containing the sensory TC nuclei LGd, VPM, VPL, and other non-TC structures (ZI, RT, and SubG). (Bi–iii). Magnifications illustrating the dominant presence of SOX14⁺GAD1⁺ interneurons in the sensory FO nuclei. (C) Anterior plane containing the VAL, centromedial (CM), anteroventral (AV), LD, and MD. The prethalamic RT is recognisable as an entirely SOX14⁺GAD1⁺ structure. (Ci). Magnification of an area of the LD containing comparable densities of SOX14⁺ and SOX14⁻ interneurons. (Cii). Magnification of an area of the MD containing SOX14⁺ and SOX14⁻ interneurons. (D) Fraction of SOX14⁺GAD1⁺ (yellow) and SOX14⁻GAD1⁺ (red) interneurons in selected TC nuclei. Above each bar the total cell counts from nine regions of interest (ROI) measuring 263 μ m by 263 μ m per each TC nucleus in three age-matched brains (three ROI per TC nucleus per brain). The average fraction of SOX14⁺GAD1⁺ interneurons deviates significantly from background level in the MD and LD. Scale bars: low magnification; overviews: \sim 1 mm; magnified areas: \sim 0.2 mm.

and multimodal associative medial, lateral, inferior, and anterior pulvinar subdivisions (MPul, LPul, IPul, and APul, respectively), FO somatosensory VPM and VPL, HO sensory PO, and FO auditory (MGv) and HO auditory (MGd) as well as non-specific HO MD and LD. With the exception of the MD and LD, all TC nuclei tested contained mostly SOX14⁺GAD1⁺ cells ($90.2 \pm 0.9\%$; mean \pm SEM; dotted line in **Figure 4D**). This may in fact be an underestimate due to the observed lower efficiency of

the *SOX14* probe compared to the *GAD1*. Analysis of the frequency distribution of the two cell classes across the tested nuclei reveals the MD and LD as outliers of an otherwise normal distribution (Motulsky and Brown, 2006). Indeed, in the MD and LD *SOX14*⁺*GAD1*⁺ cells account for $28.7 \pm 6.7\%$ and $52.3 \pm 2.8\%$ (mean \pm SEM) of the total *GAD1*⁺, respectively (Figure 4D). The presence of a sizeable population of *SOX14*⁺*GAD1*⁺ cells in the MD and LD is intriguing for their reminiscence of the *Sox14*⁺*Gad1*⁺ interneurons of the mouse (Figure 2C,D) which are also most abundant in these two non-specific HO nuclei.

We find the presence in both the mouse and marmoset of an abundant *SOX14*⁺*GAD1*⁺ interneuron class and the relative distribution of *SOX14*⁺*GAD1*⁺ and *GAD1*⁺ single positive interneurons compatible with a conserved basic organisation of interneuron diversity in rodents and primates alike.

All *Sox14*-expressing thalamic interneurons are born in the midbrain

Given the known role of *Sox14* in specifying subcortical inhibitory classes (Achim et al., 2013; Delogu et al., 2012; Guo and Li, 2019; Prekop et al., 2018; Sellers et al., 2014) and following our identification of *Sox14* as a conserved genetic marker for the larger cohort of thalamic interneurons in both the primate and rodent brain, we investigated the requirement for this gene in interneurons across TC modalities and hierarchy using the *Sox14*^{GFP/GFP} (*Sox14* knockout, KO) mouse. We have previously shown that in the *Sox14* KO there is a >90% reduction in the number of interneurons in the LGd (Jager et al., 2016). We find a comparable reduction in the number of GFP⁺ interneurons overall across the LGd, LP, VP, PO, and MG in the *Sox14* KO ($90.5 \pm 1.5\%$, $p=2.7 \times 10^{-4}$, two-sample two-tailed t-test, $n = 3$ brains/genotype; Figure 5A,B). Conversely, there was no significant change in the number of *Sox14*⁺*Gad1*⁺ cells ($p=0.4$, two-sample two-tailed t-test; Figure 5A,B) and in their distribution across TC regions (Figure 5A,C; $p>0.05$, chi-squared test, $n = 3$ brains/

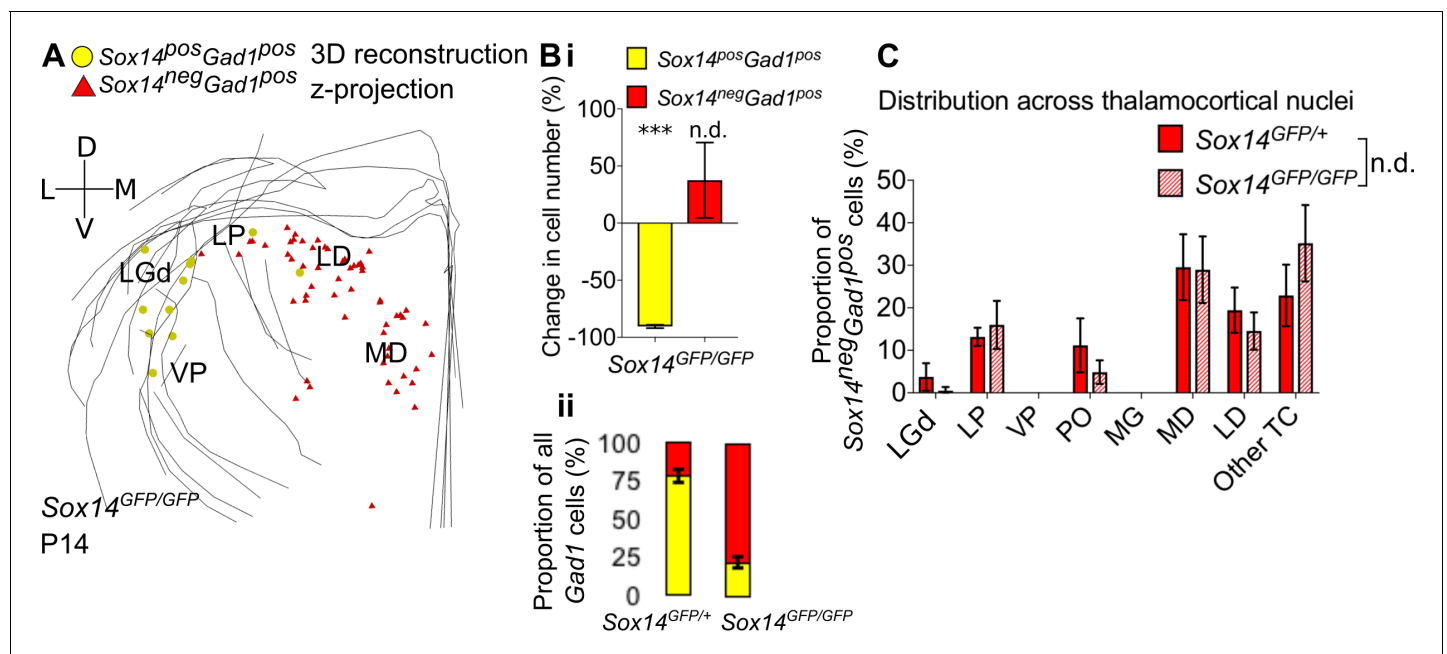


Figure 5. Differential requirement for *Sox14* highlights two distinct developmental classes. (A) Differential requirement for *Sox14* highlights two distinct developmental classes. 3D reconstruction of a representative P14 *Sox14*^{GFP/GFP} thalamus from tracing every tenth 20- μ m-thick coronal section, displayed as a z-projection and showing distribution of *Sox14*⁺*Gad1*⁺ (yellow) and *Sox14*⁺*Gad1*⁺ cells (red). (B) (i) Relative change in the number of GFP⁺*Gad1*⁺ and GFP⁺*Gad1*⁺ cells across TC regions in P14 *Sox14*^{GFP/GFP} relative to P14 *Sox14*^{GFP/+} data (mean \pm SEM, $n = 3$ brains/genotype). There is a significant reduction in the GFP⁺*Gad1*⁺ population ($p=2.7 \times 10^{-4}$, two-sample two-tailed t-test), but no statistically significant difference in the size of the GFP⁺*Gad1*⁺ group ($p=0.4$, two-sample two-tailed t-test). (ii) Proportion of GFP⁺*Gad1*⁺ cells within the total GABAergic population is decreased in the *Sox14*^{GFP/GFP} (mean \pm SEM, $n = 3$ brains/genotype). (C) Distribution of GFP⁺*Gad1*⁺ cells across TC nuclei in the *Sox14*^{GFP/+} and *Sox14*^{GFP/GFP} brains at P14 (mean \pm SEM; $n = 3$ brains/genotype). GFP⁺*Gad1*⁺ distribution is unaltered in the *Sox14* KO ($p>0.05$, chi-squared test).

genotype). These results therefore indicate that the two TC interneuron populations may already be segregated during development and represent two distinct GABAergic lineages.

LGd interneurons in the mouse derive from the midbrain (Jager et al., 2016). To explore how the molecular and spatial organisation of thalamic interneurons is generated during development more conclusively, we fate-mapped midbrain lineages and checked for their presence, distribution, and inhibitory profile across the thalamus. We crossed *En1^{Cre}* (Kimmel et al., 2000) with a *Rosa26^{Isl-GFP}* (Sousa et al., 2009) reporter line (Figure 6A; see also Table 1), as the *En1* TF gene is expressed in the midbrain and rostral hindbrain progenitors, but not in the forebrain (Sgaier et al., 2007). Analysis of the thalamus at P21 reveals GFP⁺ cells (*En1*⁺ lineage) distributed across the LGd and co-expressing GABA (Figure 6B), therefore independently validating our previous observation (Jager et al., 2016). However, like the *Sox14⁺Gad1⁺* neurons, *En1*⁺ cells were observed beyond the LGd – in the LP, VP, PO, and MG, where they were also positive for GABA (Figure 6B,C). Plotting their distribution confirmed that it is equivalent to the distribution of *Sox14*⁺ INs ($p > 0.05$, chi-squared test; Figure 6C,D). Occasional GFP⁺ cells with glia-like morphology were also observed in the thalamus. These cells were GABA[−] and were not included in any of the analyses.

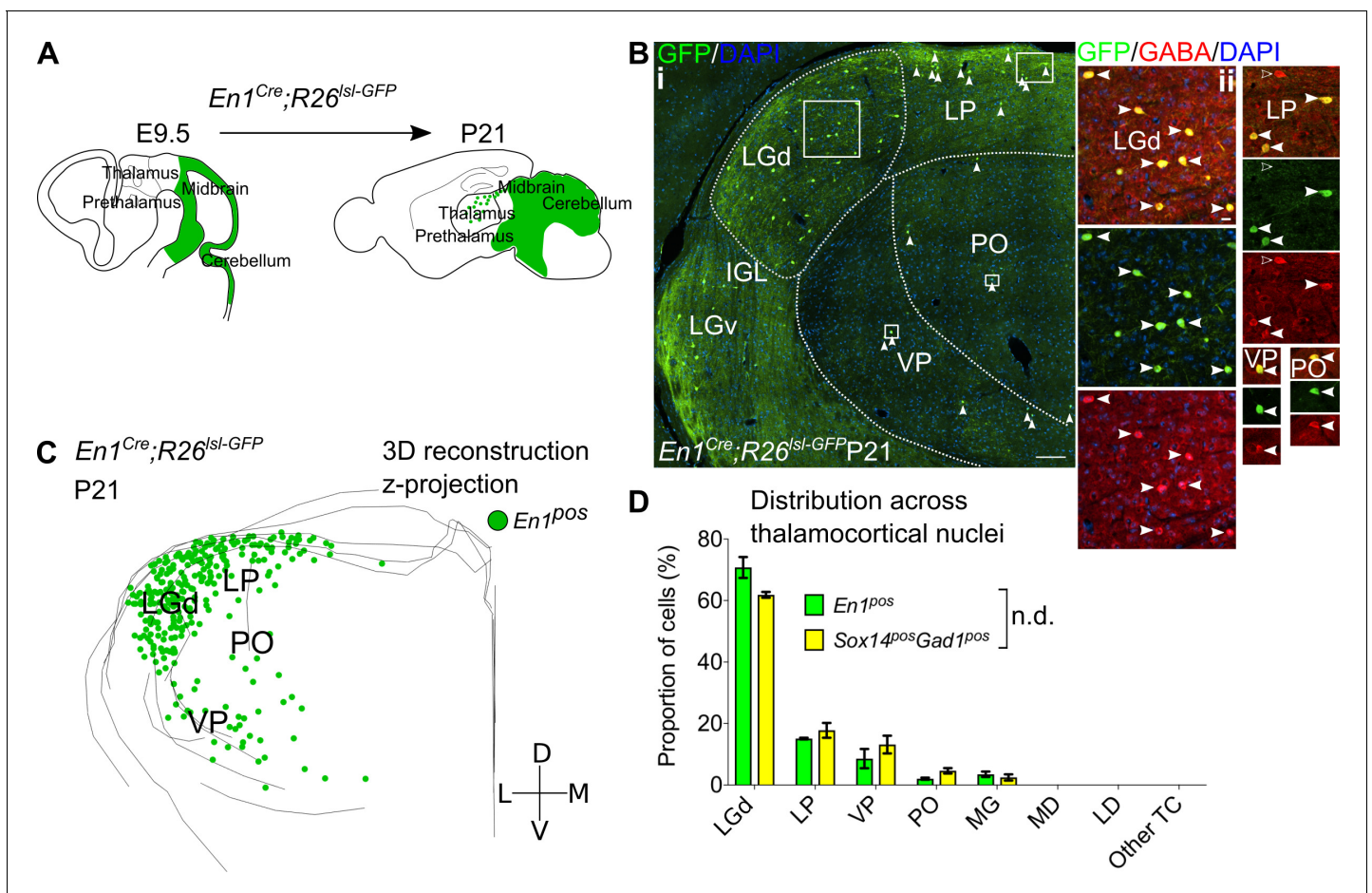


Figure 6. *Sox14*⁺ interneurons in TC regions derive from the midbrain. (A) Schematic of the fate mapping experiment: crossing *En1^{Cre}* with *Rosa26^{Isl-GFP}* reporter line permanently labels all midbrain born cells with GFP expression. (B) (i) Representative coronal section of P21 *En1^{Cre}; Rosa26^{Isl-GFP}* thalamus with *En1*⁺ cells observed in the LGd, LP, VP, and PO (considering TC regions only). For clarity some of the *En1*⁺ cells are indicated with white arrows. Scale bar, 100 μ m. (ii) *En1*⁺ cells in these regions co-express GABA (filled white arrows). Empty arrows mark GABA single-positive cells. Scale bar, 10 μ m. (C) 3D reconstruction of a representative P21 *En1^{Cre}; Rosa26^{Isl-GFP}* thalamus from tracing every sixth 60- μ m-thick coronal section, displayed as a z-projection and showing distribution of *En1*⁺ cells. (D) Distribution of *Sox14⁺Gad1⁺* and *En1*⁺ cells across TC nuclei in *Sox14^{GFP/+}* and *En1^{Cre}; Rosa26^{Isl-GFP}* brains, respectively, plotted as proportion of all the cells within each group (mean \pm SEM; $n = 3$ brains/genotype). The two populations are not differently distributed ($p > 0.05$, chi-squared test).

We therefore conclude that the *Sox14*⁺ thalamic interneurons across FO and HO TC nuclei all derive from the midbrain, and simultaneously that the *Sox14*⁺ GABAergic cells do not; the two classes thus represent distinct inhibitory lineages in TC regions, further supporting their definition as two distinct thalamic interneuron classes.

Midbrain-derived interneurons migrate along two streams into the sensory thalamus during the first postnatal week

En1^{Cre}; *Rosa26*^{Isl-GFP} line was then used to investigate the timeline and spatial trajectories of the *Sox14*⁺ interneuron precursors migrating from the midbrain into the FO and HO sensory TC regions (Figure 7A). Previously, LGd interneurons were found to populate this nucleus in the first postnatal week (Golding et al., 2014; Jager et al., 2016). We therefore looked at the numbers and migratory morphology of GFP⁺ (i.e. *En1*⁺) cells in the thalamus at E16.5, E17.5, P0.5, P1.5, and P2.5. We focused on the LGd, LP, and VP, but left out the PO and MG, due to low overall numbers of interneurons in these two regions (Figure 1D and Figure 2D).

At E16.5 no GFP⁺ cells were present in the thalamus. From E17.5 to P2.5 their numbers progressively increased in all of the regions analysed (Figure 7A,B). The number of GFP⁺ cells in the LGd at P2.5 matched previous independent reports (Golding et al., 2014), validating our counting method. Midbrain-derived interneurons therefore populate the different TC regions following a similar

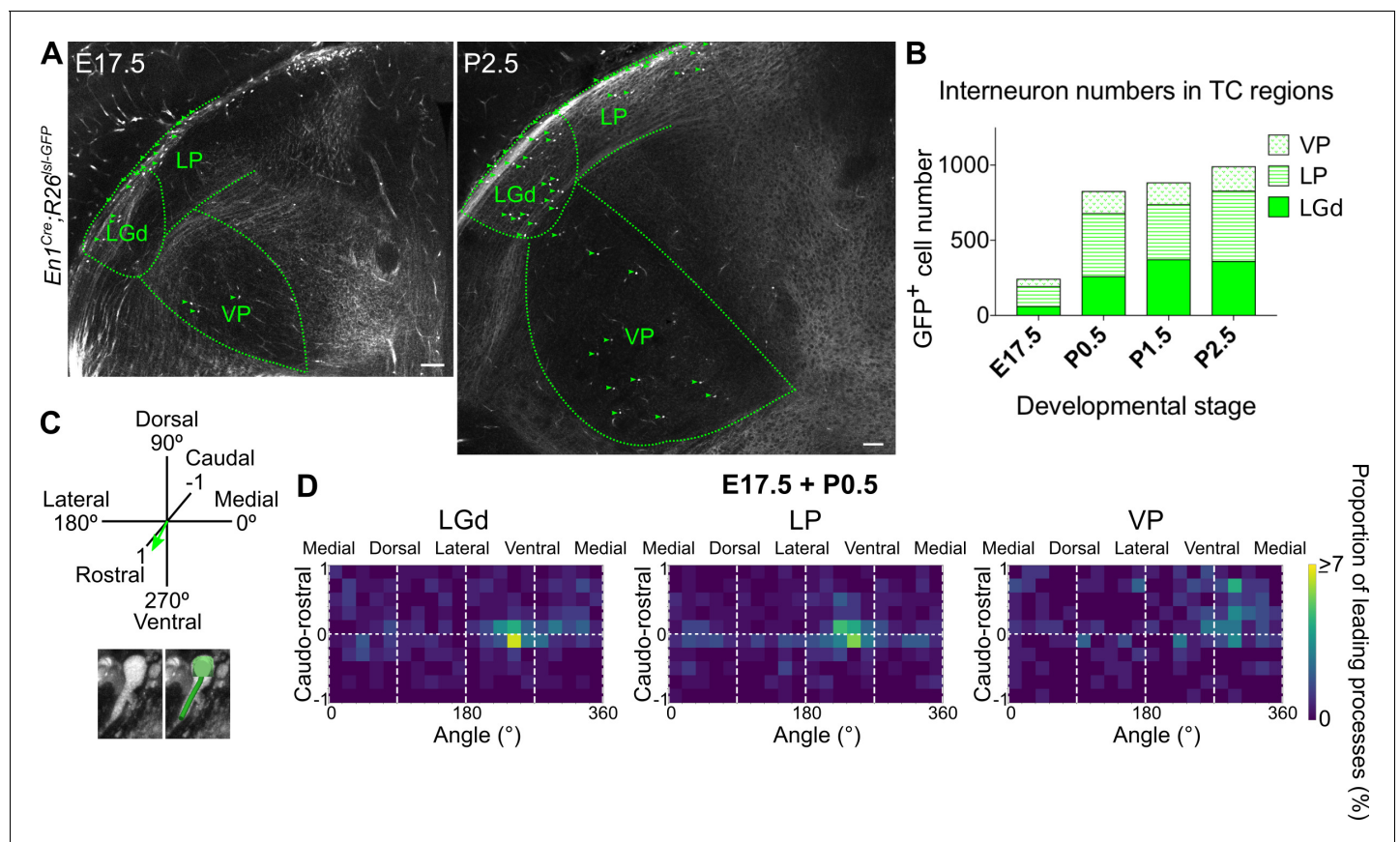


Figure 7. Midbrain-derived interneuron precursors progressively populate the thalamus from E17.5 onwards. (A) Representative coronal sections of *En1*^{Cre}; *Rosa26*^{Isl-GFP} thalamus at E17.5 and P2.5. Green arrows mark some of the GFP⁺ cells. Scale bars, 100 μm. (B) Number of GFP⁺ cells counted in the LGd, LP, and VP from E17.5 to P2.5 (mean, n = 3 brains). (C) Leading process orientation of GFP⁺ cells was determined along the caudo-rostral, ventro-dorsal, and latero-medial dimensions. (D) Frequency distribution of leading process orientation for GFP⁺ cells in the LGd, LP, and VP at E17.5 and P0.5 combined, represented in heat maps (n = 3 brains/developmental stage).

The online version of this article includes the following figure supplement(s) for figure 7:

Figure supplement 1. Distinct leading process orientation between interneuron precursors in the LGd and LP compared to VP.

timeline. Interestingly, they appear in two ventrally located nuclei (i.e. LGd and VP) simultaneously (**Figure 7A,B**), implying they use distinct routes to reach them.

To assess quantitatively the direction of migration of the larger, midbrain-derived interneuron class, we determined the leading process orientation of migrating GFP⁺ cells along all three dimensions: ventro-dorsal, latero-medial, and caudo-rostral; **Figure 7C**; (Jager et al., 2016; Paredes et al., 2016). This was plotted at a population level as frequency distribution using heat maps, for each nucleus individually, for E17.5 and P0.5 (**Figure 7D**; **Figure 7—figure supplement 1B**), as the relative increase in GFP⁺ cell numbers was the greatest between these two time points (**Figure 7B**). Moreover, there was a progressive decrease across developmental stages in the proportion of GFP⁺ cells for which migratory morphology could be identified (**Figure 7—figure supplement 1A**).

Heat maps indicate that at a population level (integrated across dimensions), GFP⁺ cells migrate into the LGd, LP, and VP in a caudo-rostral and dorso-ventral direction (**Figure 7D**), consistent with the position of the thalamus in the brain relative to their midbrain origin. However, GFP⁺ precursors in the LGd and LP have a dominant medio-lateral orientation, while those in the VP an opposite, latero-medial orientation, as can also be seen from polar histograms (**Figure 7—figure supplement 1C**). This suggests that midbrain-derived interneuron precursors enter TC regions simultaneously in two distinct streams, one migrating rostro-ventro-laterally to the LGd and LP, and the other rostro-ventro-medially to the VP, indicating a split between visual (LGd, LP) and somatosensory (VP) TC nuclei.

Sox14-negative thalamic interneurons populating HO nuclei are born in the forebrain

With the aim of identifying the origin of the Sox14[−] interneuron class in the mouse, we first looked for a positive molecular marker that would complement the absence of Sox14 expression. We made use of DropViz data [available from: dropviz.org; (Saunders et al., 2018)] and observed that within inhibitory clusters from the diencephalon, Sox14 and Pvalb show largely non-overlapping expression. It is known that Pvalb is expressed by the prethalamic RT (Clemente-Perez et al., 2017) and by telencephalic interneuron subtypes derived from the GE (Marín and Rubenstein, 2001; Tasic et al., 2016; Tremblay et al., 2016).

We therefore checked whether any thalamic interneurons are fluorescently labelled using a Pvalb-Cre mouse (Hippenmeyer et al., 2005) crossed to the conditional reporter Sun1sfGFP (Rosa26^{Isl-nuclearGFP}; see also **Table 1**). These mice contain a CAG promoter driving expression of the mouse nuclear membrane protein SUN1 (Sad1 and UNC84 domain containing 1) fused to two copies of superfolder GFP (sfGFP) inserted in the Rosa26 locus (Mo et al., 2015). Indeed, at P14 GFP was detected at the nuclear membrane of GABA-expressing neurons within the same regions populated by the Sox14[−] interneurons (**Figure 8—figure supplement 1A**), including the MD and LD, but absent from TC nuclei populated exclusively by Sox14⁺ interneurons, such as the LGd and VP (**Figure 8—figure supplement 1Ai**).

At later ages (P56) Pvalb is widely expressed in the mouse thalamus and is observed in high-density gradients in several TC nuclei [2004 Allen Institute for Brain Science. Allen Mouse Brain Atlas. Available from: mouse.brain-map.org; (Lein et al., 2007)]. Pvalb expression is not conserved across rodents and primates and cannot assist in comparative studies (Rausell and Jones, 1991). Importantly, however, in the P14 mouse 93.9% of Pvalb⁺ cells in TC regions co-expressed GABA (n = 2 brains, **Figure 8—figure supplement 1Aii, B**) and none were labelled by the En1^{Cre} reporter (0 out of 491 Pvalb⁺ cells, n = 4 En1cre; Rosa26^{Isl-GFP} mice) or Sox14^{GFP/+} reporter (0 out of 206 Pvalb⁺ cells, n = 2 Sox14^{GFP/+} mice). Therefore, we define the mouse Sox14[−] GABAergic cells as Pvalb⁺.

We found the presence of a minor population of Pvalb⁺ interneurons in the anterior thalamus of the mouse intriguing. Spatial proximity to the RT and shared marker expression (Pvalb) may suggest a prethalamic origin for the Sox14[−]Pvalb⁺ interneurons. On the other hand, progenitor domains of the telencephalic medial ganglionic eminences (MGE) and preoptic area (POA) also generate Pvalb⁺ interneurons, which are known to integrate in neocortical and hippocampal circuitries (Gelman et al., 2011; Lavdas et al., 1999; Wichterle et al., 2001; Xu et al., 2004; Xu et al., 2008), but could potentially reach the thalamus. In addition, in humans the DLX1/2/5⁺ GE generate thalamic interneurons selectively for the HO MD and pulvinar nuclei (Letinić and Kostović, 1997; Letinic and Rakic, 2001; Rakić and Sidman, 1969).

We set out first to validate the distinct origin of the Sox14⁺Pvalb⁺ interneurons by fate-mapping rostral forebrain inhibitory progenitor domains using the *Dlx5/6^{Cre}* (Monory et al., 2006) crossed to *Rosa26^{Isl-GFP}* line (Figure 8A; see also Table 1). Expression of the distal less homeobox genes *Dlx1/2/5/6* is a common ontogenetic step for all GABAergic lineages in the subpallium, prethalamus, and hypothalamus (Anderson et al., 1997a; Anderson et al., 1997b; Bulfone et al., 1993; Le et al., 2017; Liu et al., 1997; Long et al., 2009a; Long et al., 2007; Long et al., 2009b; Simeone et al., 1994; Wang et al., 2010; Yun et al., 2002).

At P14 virtually all TC Pvalb⁺ cells are a *Dlx5* lineage (GFP⁺; 99.71 ± 0.29%, n = 3 brains; Figure 8B,K) and majority of them co-expressed detectable GABA (93.6 ± 3.7%; Figure 8B,L), in line with observations from the *Pvalb^{Cre}; Rosa26^{Isl-nuclearGFP}* line (Figure 8—figure supplement 1Aii, B). We mapped the distribution of Pvalb⁺*Dlx5*⁺ cells across TC regions (Figure 8C,D) and observed that it closely recapitulates the distribution of Sox14⁺Gad1⁺ cells (Figure 8C; p>0.05, chi-squared test).

We also observed other Pvalb⁺*Dlx5*⁺ cells in the thalamus, the majority of which had a glia-like morphology and did not express GABA (Figure 8—figure supplement 1C). Occasional Pvalb⁺GABA⁺*Dlx5*⁺ cells with neuronal-like morphology were also seen (Figure 8—figure supplement 1C,D), suggesting leaky Cre activity in some cases. That all Pvalb⁺ cells in TC nuclei are labelled with GFP argues against this being an artefact of leaky reporting. Pvalb⁺GABA⁺*Dlx5*⁺ cells were not considered in any of the analyses.

While the *Dlx5/6^{Cre}* fate mapping confirmed the Sox14⁺Pvalb⁺ cells as a distinct, forebrain-derived interneuron in the mouse thalamus, it does not resolve between the different rostral forebrain GABAergic territories of the telencephalon and diencephalon, the latter comprised of the prethalamus and the hypothalamus.

To further discriminate between telencephalic and prethalamal/hypothalamic GABAergic progenitors, we took advantage of the restricted expression of the Forkhead box gene *Foxd1* during the neurogenic period in the prethalamus and hypothalamus (Newman et al., 2018; Shimogori et al., 2010) and fate-mapped *Foxd1*⁺ lineages by crossing the *Foxd1^{Cre}* (Humphreys et al., 2008) with the *Rosa26^{Isl-nuclearGFP}* reporter line, described earlier (Figure 8E; see also Table 1). At P14, GFP was readily detectable at the nuclear membrane of neurons in the prethalamus and, as previously reported (Newman et al., 2018), in non-neuronal endothelial cells of the brain's vascular system. Within TC nuclei, colabelling with Pvalb and GABA revealed that the vast majority of thalamic Pvalb⁺ cells derive from *Foxd1*⁺ progenitors (85.56 ± 2.1%; 297 Pvalb⁺ cells from n = 5 *Foxd1^{Cre}; Rosa26^{Isl-nuclearGFP}* mice) and that virtually all Pvalb⁺*Foxd1*⁺ cells contained detectable GABA (96.43 ± 1.63%). Other GFP nuclei that are neither Pvalb⁺ nor GABA⁺ likely belong to endothelial cells in blood vessels and were NeuN negative (data not shown). The minor residual fraction of Pvalb⁺GFP[−] cells is similarly distributed across TC nuclei and likely to reflect incomplete loxp recombination or very low expression at the *Rosa26* locus.

The fate mapping with *Foxd1^{Cre}* is therefore consistent with a rostral diencephalic origin of the Sox14⁺Pvalb⁺ interneurons, rather than a telencephalic one, but cannot resolve between prethalamus and hypothalamus. However, *Dlx5*⁺ GABAergic domains of the posterior and ventral hypothalamus can be defined by partially overlapping expression of *Lhx6* or *Nkx2.1* (Díaz et al., 2014; Shimogori et al., 2010), two transcription factor genes that are also expressed in the GE, but not expressed in the prethalamus (Figure 8G,I). Therefore, we tested for a potential hypothalamic origin of the Sox14⁺Pvalb⁺ interneurons by fate mapping GABAergic subdomains of the hypothalamus using the *Nkx2.1^{Cre}* (Xu et al., 2008) and the *Lhx6^{Cre}* (Fogarty et al., 2007) line crossed to the *Rosa26^{Isl-GFP}* reporter (Table 1).

The *Dlx5*⁺*Nkx2.1*⁺ hypothalamic GABAergic progenitors are located in the tuberal and retrotuberal area of the basal hypothalamus (Díaz et al., 2014; Morales-Delgado et al., 2014; Shimogori et al., 2010). Hence, we fate-mapped *Nkx2.1*⁺ lineages with the *Nkx2.1^{Cre}* line (Figure 8G) and investigated the presence of GFP⁺Pvalb⁺ co-expressing neurons in TC regions, at P14. While GFP⁺ cells are present in thalamic territory, none of the TC Pvalb⁺ cells belonged to a *Nkx2.1*⁺ lineage (GFP⁺Pvalb⁺0%, n = 3 brains; Figure 8Hi,Hii,K), therefore excluding the postero-ventral hypothalamus as a possible source of the Pvalb⁺ thalamic interneurons. Importantly, *Nkx2.1* is also a well-established marker of GABAergic progenitors in the MGE and POA (Shimamura et al., 1995; Sussel et al., 1999) and defines several cortical interneuron lineages including the fast spiking Pvalb⁺ interneurons, the somatostatin⁺ interneurons (Fogarty et al., 2007; Pleasure et al., 2000;

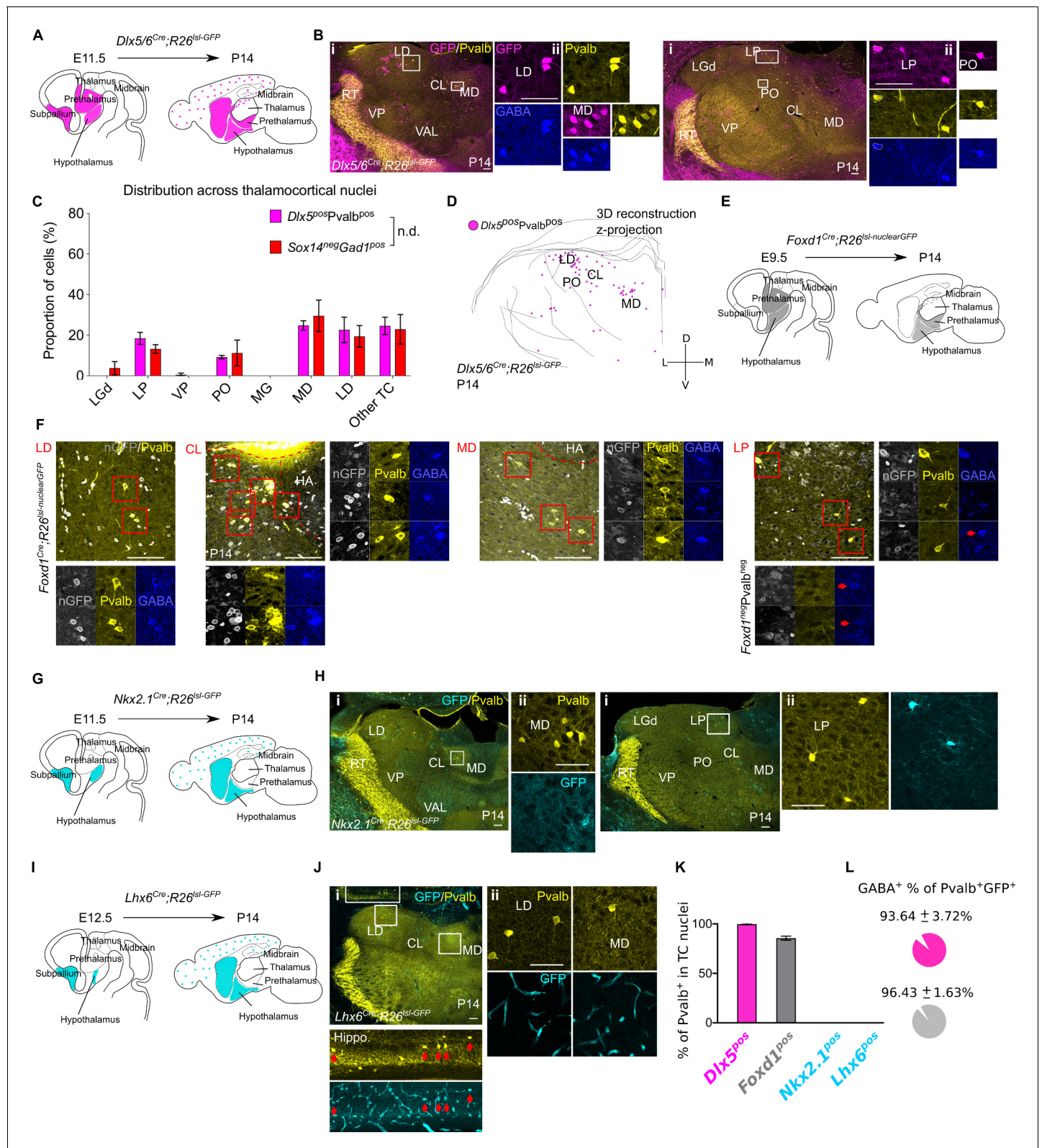


Figure 8. *Sox14⁺Pvalb⁺* interneurons in TC regions derive from the *Dlx5^{pos}*, *Foxd1^{pos}*, *Nkx2.1^{neg}*, and *Lhx6^{neg}* rostral forebrain. (A) Schematic of the fate mapping experiment: crossing *Dlx5/6^{Cre}* with *Rosa26^{Isl-GFP}* reporter line permanently labels all ventral telencephalic, hypothalamic, and prethalamic-born cells with GFP expression. (B) (i) Representative coronal sections of P14 *Dlx5/6^{Cre}; Rosa26^{Isl-GFP}* thalamus with *Dlx5^{pos}Pvalb^{pos}* cells present in the MD, LD, CL, VAL, VM, LP, and PO (considering TC regions only). Scale bar, 100 μ m. (ii) *Dlx5^{pos}Pvalb^{pos}* cells in TC regions co-express GABA. Scale bar, 100 μ m. Figure 8 continued on next page

Figure 8 continued

(C) Distribution of $Dlx5^+Pvalb^+$ and $Sox14^+Gad1^+$ cells across TC nuclei in P14 $Dlx5/6^{Cre}; Rosa26^{lsI-GFP}$ and $Sox14^{GFP/+}$ brains, respectively, plotted as proportion of all the cells within each group (mean \pm SEM, $n = 3$ brains/genotype). The two populations are not differently distributed ($p > 0.05$, chi-squared test). (D) 3D reconstruction of a representative P14 $Dlx5/6^{Cre}; Rosa26^{lsI-GFP}$ thalamus from tracing every sixth 60- μ m-thick coronal section, displayed as a z-projection and showing distribution of $Dlx5^+Pvalb^+$ cells. (E) Schematic of the fate mapping experiment crossing the $Foxd1^{Cre}$ with $Rosa26^{lsI-nuclearGFP}$ reporter line to permanently label hypothalamic and prethalamic-born cells with nuclear membrane localised GFP. (F) Representative coronal sections of P14 $Foxd1^{Cre}; Rosa26^{lsI-nuclearGFP}$ thalamus with $Foxd1^+Pvalb^+$ cells present in the LD, CL, MD, and LP. Scale bar, 100 μ m. Enlarged areas (red boxes) showing $Foxd1^+Pvalb^+$ cells co-expressing GABA. Additional GABA $^+$ but $Foxd1^-Pvalb^-$ neurons are also visible in the LP (red arrows). HA: habenula. (G) Schematic of the fate mapping experiment: crossing $Nkx2.1^{Cre}$ with $Rosa26^{lsI-GFP}$ reporter line permanently labels some hypothalamic and all MGE-born cells with GFP expression. (H) (i) Representative coronal sections of P14 $Nkx2.1^{Cre}; Rosa26^{lsI-GFP}$ thalamus with $Pvalb^+$ and $Nkx2.1^+$ cells present in the MD, LD, CL, VAL, VM, LP, and PO (considering TC regions only). Scale bar, 100 μ m. (ii) $Nkx2.1^+$ cells in TC regions do not co-express $Pvalb^+$. Scale bar, 100 μ m. (I). Schematic of the fate mapping experiment crossing the $Lhx6^{Cre}$ with $Rosa26^{lsI-GFP}$ reporter line to permanently label some hypothalamic and MGE born cells with GFP. (J) (i) Representative coronal section of P14 $Lhx6^{Cre}; Rosa26^{lsI-GFP}$ showing GFP $^+Pvalb^+$ cells in the hippocampus (red arrows) and GFP $^+Pvalb^+$ present in the MD, LD, and CL nuclei of the thalamus (considering TC regions only). Scale bar, 100 μ m. (ii) GFP $^+$ cells in TC regions are endothelial and do not co-express $Pvalb^+$. Scale bar, 100 μ m. (K) Proportion of $Pvalb^+$ cells in TC regions that are $Dlx5^+$, $Foxd1^+$, $Nkx2.1^+$, or $Lhx6^+$ at P14 (mean \pm SEM, $Dlx5^+ n = 3$ brains, $Foxd1^+ n = 5$ brains, $Nkx2.1^+ n = 3$ brains, and $Lhx6^+ n = 4$ brains). (L) Proportion of $Dlx5^+Pvalb^+$ and $Foxd1^+Pvalb^+$ cells in TC regions co-expressing GABA at P14 (mean \pm SEM, $Dlx5^+ n = 3$ brains, $Foxd1^+ n = 5$ brains). The online version of this article includes the following figure supplement(s) for figure 8:

Figure supplement 1. $Pvalb$ is a marker for $Sox14^+$ thalamic interneurons.

Xu et al., 2004; Xu et al., 2008), and from the POA, Npy^+ , or $Reelin^+$ interneurons (*Gelman et al., 2011; Gelman et al., 2009*). Hence this result strengthens the evidence against a telencephalic contribution to thalamic interneurons in the mouse.

Although we did not conduct a detailed investigation of $Pvalb^-$ cell types labelled by the $Nkx2.1^{Cre}; Rosa26^{lsI-GFP}$ reporter in the thalamus, we noted several glia-like morphologies that were also negative for the pan-neuronal marker NeuN (data not shown).

The hypothalamic domain of $Lhx6$ expression only partially overlaps with that one of $Nkx2.1$ and defines additional $Dlx5^+$ GABAergic progenitors of the hypothalamus (*Díaz et al., 2014; Kim et al., 2020; Shimogori et al., 2010*). Hence, we fate-mapped $Lhx6^+$ lineages with the $Lhx6^{Cre}$ line (*Figure 8I*) and investigated the presence of GFP $^+Pvalb^+$ co-expressing neurons in TC regions, at P14. As no GFP $^+$ cells were present in thalamic territory (GFP $^+Pvalb^+$ 0%, $n = 4$ brains; *Figure 8Ji, Jii, K*), we excluded the alar hypothalamus as a potential source of the $Pvalb^+$ thalamic interneurons. Importantly, $Lhx6$ marks also the vast majority of GE-derived telencephalic interneurons (*Zhao et al., 2008*), hence this result further confirms that the telencephalic GE are an unlikely source of thalamic interneurons in the mouse.

Altogether, we therefore conclude that the rarer $Sox14^+$ thalamic interneuron class is a distinct lineage compared to the larger, midbrain-born $Sox14^+$ thalamic interneuron class, and that originates from $Dlx5^+Foxd1^+$ progenitors in the prethalamus.

Discussion

Our study reveals a previously unappreciated complexity of GABAergic interneurons in the mouse TC nuclei, demonstrating that interneurons are not restricted to the FO visual thalamus, but present across modalities and hierarchical levels, including limbic structures.

We recognise two broad thalamic interneuron classes, defined by their origin in either the $En1^+$ midbrain or the $Dlx5^+Foxd1^+Nkx2.1^-Lhx6^-$ rostral forebrain. However, the two ontogenetic programmes contribute differentially to interneuron numbers, with the midbrain-derived class overwhelmingly more abundant.

The midbrain-derived interneurons depend on the $Sox14$ transcription factor, a gene that we had previously implicated in LGd interneuron differentiation (*Jager et al., 2016*) and a known postmitotic marker for GABAergic subtype neurogenesis in the brainstem (*Achim et al., 2013; Achim et al., 2014; Huisman et al., 2019; Prekop et al., 2018*). Taking advantage of a $Sox14^{GFP}$ mouse line, we now provide absolute numbers and standardised anatomical distribution of this major class of interneurons across the entire thalamus, in the $Sox14^{GFP/+}$ C57Bl/6 genetic background.

Rather than representing a peculiarity of mice, the midbrain ontogenetic programme may be the dominant source of thalamic interneurons in larger-brained mammals, as suggested by the

identification of $SOX14^+GAD1^+$ interneurons across virtually all TC nuclei in the interneuron-rich thalamus of the marmoset. This finding is consistent with the recent report of molecularly defined classes of GABAergic interneurons in the macaque and human thalamus, where $SOX14$ expression differentiates between interneuron subclasses. Interestingly, $SOX14^+$ interneurons are $DLX1-6^+$, in agreement with the ontogenetic plan that we propose here (Bakken et al., 2020). Consistent with a conserved midbrain ontogeny of thalamic interneurons, Jones previously described late appearance of interneurons in the ferret and macaque thalamus, progressively from caudal towards rostral nuclei (Hayes et al., 2003; Jones, 2002). It can also be seen from the BrainSpan Atlas of the Developing Human Brain [BrainSpan Atlas of the Developing Human Brain; available from: <http://www.brain-span.org>; (Miller et al., 2014)] that both $GAD1$ and $SOX14$ expression increase in the dorsal thalamus in the mid-prenatal period (from postconception week 16), which is also consistent with a migration of midbrain-born interneurons into these regions.

Interestingly, grafting experiments using chick and quail embryos demonstrated a potential for midbrain cells to populate retino-recipient nuclei in the chick diencephalon (Martinez and Alvarado-Mallart, 1989). The grafted midbrain cells were observed migrating tangentially at the surface of the diencephalon and seemingly through the host optic tract before invading the regions targeted by the retinal projections (Martinez and Alvarado-Mallart, 1989). The neurotransmitter identity of these migrating cells is unknown, but their midbrain origin and distribution across the thalamus resemble the mouse $Sox14^+$ interneurons, suggesting that in birds too, the largest cohort of interneurons is a midbrain lineage. Relatedly, lineage tracing in chick, using a retroviral library, indicated that clonally related siblings can populate both the diencephalon and mesencephalon (Golden and Cepko, 1996), in keeping with a revised model of evolutionary relationship of caudal diencephalon and midbrain (Albuxech-Crespo et al., 2017). The distribution of $Sox14^+$ interneurons observed in the mouse is similar to the one described in the opossum, a living marsupial thought to resemble early mammals in the fossil record (Penny et al., 1984). Intriguingly, rather than spreading throughout the nucleus, interneurons occupy the lateral subdivision of the LP in the adult opossum thalamus (Penny et al., 1984), reminiscent of the route taken by migrating midbrain-derived interneuron precursors in the developing mouse thalamus.

While the emerging picture points to a midbrain ontogeny for the largest fraction of thalamic interneurons, this is not sufficient to explain the overall developmental complexity of interneurons in the thalamus. In both the mouse and marmoset, we now report the presence of $Sox14^+Gad1^+$ interneurons with drastically more restricted distribution. In the mouse this interneuron class is found enriched in HO TC nuclei (the non-specific MD and LD, but also sensory-related LP and PO). Similarly, in the marmoset $SOX14^+GAD1^+$ interneurons are also a minor class enriched in the HO nuclei MD and LD in the anterior portion of the thalamus. This specific distribution is intriguing as it may reflect the requirement in some associative nuclei for interneurons with unique functional properties that the larger midbrain-derived class cannot provide. Such hypothetical evolutionary drive is not dissimilar to the one previously proposed for some interneurons of the human thalamus, where GE-derived $DLX1/2^+$ interneurons were shown to migrate into associative nuclei MD and pulvinar (Letinić and Kostović, 1997; Letinić and Rakic, 2001; Rakic and Sidman, 1969).

While a dual midbrain and forebrain ontogeny of thalamic interneurons emerges as the conserved mammalian blueprint for thalamic interneuron organisation, with the midbrain-derived class contributing the largest proportion of interneurons and the forebrain-derived class enriched in selected HO TC nuclei, species-specific differences also exist. In the mouse, midbrain- and forebrain-derived interneurons are spatially segregated along clear anatomical and functional subdivisions of the thalamus, in the marmoset, presumptive midbrain-derived interneurons expanded dramatically and are more broadly distributed. The forebrain-derived interneurons are not found in precisely the same set of associative TC nuclei across mouse and marmoset, while the forebrain-derived human lineage previously described is thought to be an evolutionary innovation that migrates into the thalamus from the GE along transient anatomical structures that are not present in rodents and non-human primates (Letinić and Kostović, 1997). Consistent with this, our data support a model whereby the mouse, and by extension the marmoset $Sox14^+Gad1^+$ interneurons are specified in the $Nkx2.1^+Lhx6^+Dlx5^+Foxd1^+$ prethalamus, rather than subpallium. Technical limitations make a detailed assessment of lineage descent in non-human primates and humans more challenging and whether humans retained the prethalamic interneuron class that we described here is currently unknown.

Further investigation of species-specific differences may provide important cues to trace the evolution of the mammalian TC system, using interneurons as the key to unravel its complexity.

Materials and methods

Animals

The mice were housed in the animal facilities at King’s College London under standard conditions on a 12 hr:12 hr dark/light cycle, with unrestricted access to water and food. Housing and experimental procedures were approved by the King’s College London Ethical Committee and conformed to the regulations of the UK Home Office personal and project licences under the UK Animals (Scientific Procedures) 1986 Act. Both female and male mice were used in a randomised way across experiments. The morning when the vaginal plug was observed was designated as embryonic day (E) 0.5 and the day of birth as postnatal day (P) 0.5 (see also [Table 1](#)).

Callithrix jacchus

A total of seven New World marmoset (*C. jacchus*) monkeys were used in this study. All experiments were conducted in accordance with the guidelines approved by the RIKEN Institutional Animal Care (W2020-2-022).

Immunohistochemistry and ISH

Mice were transcardially perfused with 4% PFA and the brains dissected and postfixed in PFA at 4°C overnight, then washed in PBS for at least 24 hr at 4°C. For ISH, brains were stored in PFA for 5 days, to minimise RNA degradation, and all the subsequent solutions were treated with diethyl pyrocarbonate (DEPC; AppliChem). The brains were cryoprotected in a sucrose gradient (10–20–30%), frozen on dry ice, and cryosectioned as 20 µm coronal sections collected on Superfrost Ultra Plus slides (Thermo Scientific) for ISH, or as 60 µm free-floating coronal sections for IHC.

Immunohistochemistry

Brain sections were washed in PBS three times and blocked in 2–7% normal goat serum (NGS) solution (in 1× PBS, 0.1–0.3% Triton-X100) for 2 hr at room temperature (RT). Primary antibodies ([Table 2](#)) were diluted in blocking solution and incubated with the sections (as stated in the table). This was followed by three 30 min PBS washes, and incubation in secondary antibodies ([Table 2](#)) diluted 1:500 in blocking solution, for 2 hr at RT. After two 30 min PBS washes, the sections were incubated in DAPI for 30 min (1:40,000 dilution in PBS; Life Technologies) and mounted using Pro-Long Gold mounting media (Invitrogen).

In situ hybridisation

Gad1 antisense RNA probe was transcribed in vitro from full-length cDNA template (IMAGE ID: 5358787). The probe was diluted to a final concentration of 800 ng/ml in hybridisation buffer (50% formamide, 10% dextran sulphate, 1 mg/ml rRNA, 1× Denhardt’s solution, 0.2 M NaCl, 10 mM Tris HCl, 5 mM NaH₂PO₄·2H₂O, 1 mM Tris base, 50 mM EDTA) and applied onto the slides, which were

Table 2. Antibodies.

Antibody	Dilution	Incubation time	Source
Rabbit anti-GABA	1:2000	2X ON, 4°C	Sigma, A2052
Chicken anti-Gfp	1:5000	2X ON, 4°C	Abcam, Ab13970
Mouse anti-parvalbumin	1:2000	1X ON, 4°C	Sigma-Aldrich, P3088
Goat anti-chicken Alexa-488	1:500	2 hr, RT	Invitrogen, A11039
Goat anti-rabbit Alexa-568	1:500	2 hr, RT	Invitrogen, A11036
Goat anti-rabbit Alexa-647	1:500	2 hr, RT	Invitrogen, A21245
Goat anti-mouse Alexa-568	1:500	2 hr, RT	Invitrogen, A11004
Goat anti-mouse Alexa-635	1:500	2 hr, RT	Invitrogen, A31575

incubated in a humidified chamber at 65°C overnight. The slides were then washed three times for 30 min in wash buffer (50% formamide, 1× SSC, 0.1% Tween) at 65°C, two times for 30 min in MABT buffer (100 mM maleic acid, 150 mM NaCl, 0.1% Tween-20) at RT, and blocked for 2 hr at RT (2% Boehringer Blocking Reagent (Roche), 20% inactivated sheep serum in MABT). Sheep a-DIG alkaline phosphatase conjugated antibody (Roche, 11093274910) was diluted 1:2000 in the blocking solution and incubated with the slides overnight at 4°C. This was followed by five 20 min washes in MABT and two 20 min washes in the AP buffer (0.1M Tris-HCl pH 8.2, 0.1%-Tween-20). Fast red TR/Naphthol AS-MX tablets (Sigma) were dissolved in the AP buffer and applied onto the slides for colour reaction for 3–6 hr at RT in the dark. The slides were then washed three times for 20 min in PBS before proceeding with IHC for GFP as described above. Sox14^{GFP/+} and Sox14^{GFP/GFP} sections were always processed in parallel.

ISH in *C. jacchus*

Fluorescent ISH was performed as previously described (Watakabe *et al.*, 2006) with some modifications. Riboprobes incorporating digoxigenin (DIG) and fluorescein (FL) were hybridised overnight. After washing, FL- and DIG-labelled probes were each detected in different ways. For detection of the DIG probes, the sections were incubated with an anti-DIG antibody conjugated with horse radish peroxidase (HRP) (1/500, Roche Diagnostics) for 6 hr at room temperature. After washing in TNTx (0.1 M Tris-HCl, pH 7.5, 0.15 M NaCl, 0.05% Triton X-100) three times for 5 min, the sections were treated with 1:100 diluted TSA-Plus (DNP) reagents (Perkin Elmer) for 20 min. After washing in TNTx 3 × 10 min, the sections were incubated for 2 hr at room temperature with an anti-DNP antibody conjugated with Alexa 488 (1/500, Invitrogen). After quenching HRP activity and washing, the sections were incubated for 2 hr at room temperature with an anti-FL antibody conjugated with HRP (1/500, Roche Diagnostics) followed by reaction with TSA biotin reagents (Perkin Elmer) and visualisation with streptavidin conjugated with Alexa594 (Invitrogen).

Quantifying distribution of neuronal populations in histological sections

In mice

Confocal z-stacks covering the extent of the thalamus across all axes (caudo-rostral, ventro-dorsal, and latero-medial) were acquired using either Nikon A1R inverted confocal, inverted spinning disk Nikon Ti microscope or Olympus VS120 slide scanner, with 10× (NA 0.30 Plan Fluor DLL) and 20× (NA 0.75 Plan Apo VC or UPLSAPO NA 0.75) objectives. The stacks were then viewed with the Neurolucida software. TC nuclei were identified from the DAPI counterstain, using cytoarchitectonically recognisable structures, such as the LGd, the habenular complex, the RT, the anterior pretectum, and the fasciculus retroflexus (fr), as landmarks for orientation and reference. When the nuclear GFP reporter was used, only cells with detectable DAPI signal were included in the calculations. The cells of interest (Table 3) were assigned to TC regions by comparing the sections to the Allen Brain Reference Atlas and annotated and counted manually. For each brain, only one hemisphere was analysed (chosen in a randomised way). For experiments using *Gad1*⁺ and *Chrna6*⁺ ISH data from the Allen Mouse Brain Atlas resource [2004 Allen Institute for Brain Science. Allen Mouse Brain Atlas. Available from: mouse.brain-map.org; (Lein *et al.*, 2007)], all images of P56 C57BL/6J coronal brain sections containing the thalamus were downloaded for each gene (every eighth 25-µm-thick section, sampling every 200 µm across the thalamus), and analysed in the same way as described above.

In marmoset

Images were acquired with a fluorescence microscope BZ-X810 (Keyence) or BZ-9000 (Keyence). Representative coronal sections at anterior, intermediate, and posterior levels were analysed manually, by delineating nuclear boundaries according to the neonate Marmoset Gene Atlas, RIKEN CBS, Japan (<https://gene-atlas.brainminds.riken.jp>). Cell counting was conducted using the Cell Counter Plugin and ROI manager in ImageJ (Schindelin *et al.*, 2015). Within the boundaries of each TC nucleus analysed, three ROIs of 263 µm by 263 µm were positioned randomly and their content of single positive or double positive cells added together to generate a representative fraction of *GAD1*⁺ and *GAD1*⁺*SOX14*⁺ cells for the TC nucleus (no *SOX14* single positive cells were detected). Counts were replicated in three age matched brains to calculate mean ± SEM.

Table 3. Genetic identity of cells counted across TC regions and technical details of corresponding experiments.

Transgenic line	Age	Cells annotated/ counted	Number of brains	Sampling	Section thickness (µm)
Mouse					
Sox14 ^{GFP/+}	P21	GFP ⁺	3	Whole thalamus	50 (10; optical sections)
Sox14 ^{GFP/+}	P14	GFP ⁺ and Gad1 ⁺	3	Every tenth coronal section	20
Sox14 ^{GFP/+}	P14	GFP ⁺ and Pvalb ⁺	2	Every sixth coronal section	60
Sox14 ^{GFP/GFP}	P14	GFP ⁺ and Gad1 ⁺	3	Every tenth coronal section	20
En1 ^{Cre} ; Rosa26 ^{Isl-GFP}	P21-30	GFP ⁺	3	Every sixth coronal section	60
En1 ^{Cre} ; Rosa26 ^{Isl-GFP}	P14	GFP ⁺ and Pvalb ⁺	3	Every sixth coronal section	60
Dlx5/6 ^{Cre} ; Rosa26 ^{Isl-GFP}	P14	GFP ⁺ , Pvalb ⁺ , and GABA ⁺	3	Every sixth coronal section	60
Foxd1 ^{Cre} ; Rosa26 ^{Isl-nuclearGFP}	P14	GFP ⁺ , Pvalb ⁺ , and GABA ⁺	5	Every sixth coronal section	30
Nkx2.1 ^{Cre} ; Rosa26 ^{Isl-GFP}	P14	GFP ⁺ and Pvalb ⁺	3	Every sixth coronal section	60
Lhx6 ^{Cre} ; Rosa26 ^{Isl-GFP}	P14	GFP ⁺ and Pvalb ⁺	4	Every sixth coronal section	60
Pvalb ^{Cre} ; Rosa26 ^{Isl-nuclearGFP}	P14	GFP ⁺ , Pvalb ⁺ , and GABA ⁺	2	Every sixth coronal section	60
Marmoset					
Wild type	P0	SOX14 and GAD1	3	Representative anterior, intermediate and posterior planes	28

3D reconstructions of cell distributions from histological sections

3D reconstructions of cell distributions (Table 3) across thalamic regions were generated for each brain separately using the Neurolucida software (MBF Bioscience), from the acquired confocal z-stacks or Allen Mouse Brain Atlas ISH data as described above. For each image the outline of the thalamus and the surrounding structures were manually traced using the ‘contour’ function and the cells were annotated with the ‘marker’ function, placed at the centre of the soma. Traced images were then aligned in sequential rostro-caudal order, manually for each brain, using tissue landmarks (midline and clearly recognisable structures, for example, LGd, RT, habenula, hippocampus) for reference, and their spacing in the rostro-caudal dimension was preserved according to the sampling used for each brain.

3D reconstructions of cell distributions by whole brain serial two photon imaging

Sox14^{GFP/+} mouse brain samples (P21; n = 3) were embedded in a 4.5% oxidised-agarose solution containing agarose (type 1; Sigma), 10 mM NaIO4 (Sigma) and 50 mM phosphate buffer (PB). Samples were imaged with TissueCyte 1000 (Ragan et al., 2012) with a custom cooling system (JULABO UK Ltd.) for serial two-photon (STP) tomography across the complete mouse brain. Physical sectioning was performed every 50 µm with optical sectioning every 10 µm. A 16×, 0.8 NA immersion objective (Nikon Inc) acquired 1 × 1 mm image tiles at spatial resolution 0.54 µm with a 12 × 10 tiling mosaic required to obtain a complete coronal tissue section. Laser (Chameleon Ultra II, Coherent) excitation was conducted at 920 nm for GFP excitation with three PMT channel acquisition for red, green, and blue wavelength collection. STP imaging occurred over 5 days and generated 3.5 terabytes of data per brain. Tiled data was stitched alongside STP acquisition using a custom Python and ImageJ/Fiji pipeline.

STP data sets of each mouse brain were down-sampled to 10 µm isotropic voxel size and registered with the Allen CCF3 average atlas using Elastix (Klein et al., 2010). Registration was performed from average atlas (moving) to down-sampled STP (fixed) using a combination of rigid, affine and b-spline transformation steps, executed using a multiresolution approach for robust global and local structure registration. An advanced Mattes Mutual Information similarity metric and an

adaptive stochastic gradient descent objective optimiser were used at each transformation step with the transformation at each step combined into a final transformation map which was applied to the CCF3 annotation atlas and a custom hemisphere atlas used to distinguish structures across hemisphere. Registration resulted in a spatial mapping from the STP data to the CCF3 atlas space allowing the delineation of thousands of anatomical structures according to the Allen Brain Atlas hierarchically organised taxonomy.

For automated cell counting, a U-Net (Ronneberger O., 2015) deep learning network was trained to segment fluorescently labelled cells in STP and confocal data sets. During training, 219 images of fluorescently labelled cells (512×512 pixels; $0.54 \mu\text{m}$ voxel size) were manually segmented using ImageJ/Fiji. Images came from STP and confocal data sets of GFP labelled cells from transgenic mouse lines and viral tracing studies and contained GFP expression localised to soma and dendritic and axonal structures. During manual segmentation, only soma localised fluorescence was labelled. To increase generalisation of the network for robust performance on new data, drop out layers at 50% probability were introduced into the network, plus image augmentation was used to increase the initial 219 image data set by 56-fold. Augmentation operations included image flipping, rotation in the range -360° to $+360^\circ$, zooming in the range 90–110%, skewing, a random elastic distortion using a grid size of 10 pixel spacing, shearing and a custom Poisson noise addition. Some transformations were assisted using the Python package Augmentor (D Bloice et al., 2017), with the custom Poisson noise generation written as a class to interface with the Augmentor package. Each transformation was given a 50% probability of occurring and resulted in a final training data set of 12,264 image and annotated pairs. Training data was split 75% (9198 samples) for training the network and 25% (3066 samples) for validating the network with conscious effort to maintain class balance between the training and validation to prevent overfitting or loss issues during training.

The model was trained with the ELU activation function, using an Adam optimiser with a Binary Cross-entropy loss function. A batch size of 8 was used with early stopping evoked if a validation dice loss score did not improve after 30 epochs of training. Model training was performed on a workstation equipped with a NVIDIA Titan Xp GPU using Python and the TensorFlow 2.0 platform.

For automated thalamus counting, all structures belonging to the thalamus, according to the Allen Brain Atlas hierarchically organised taxonomy, were extracted from the registered STP data sets using masks upsampled to the original $0.54 \mu\text{m}$ data and fed into the trained network for automated segmentation. Correction for oversampling of cells in the axial axis was done by grouping detected cells if they overlapped within a radius of $10 \mu\text{m}$, and subsequently keeping the centrally positioned cell in the axial axis. Automated counting in the entire thalamus took 7 hr per sample using an Ubuntu Intel(R) Core(TM) i9-7980XE CPU at 2.60 GHz workstation, with 32 cores and 128 GB RAM.

NND calculations

Nearest neighbour distance (NND) was determined for the *Sox14⁺Gad1⁺* and *Sox14⁻Gad1⁺* cells from the 3D reconstructions of their distributions. The cells' coordinates in 3D were generated by Neurolucida and analysed using a custom Python script and the Pandas library (McKinney et al., 2010) to calculate NNDs separately for each group and between the two groups, for each *Sox14^{GFP/+}* brain individually. The data was then normalised to the largest NND within each data set (each individual group and between groups sets for each brain) averaged across the brains (mean \pm SEM) and plotted as cumulative distribution. Normalisation allows us to plot their cumulative distribution as a fraction of the maximum distance, though even before normalisation of the curves were broadly similar. Statistically significant differences between the distributions were verified using the two-sample Kolmogorov–Smirnov test, implemented in the SciPy library (Jones et al., 2001).

Migratory morphology analysis

E16.5, E17.5, P0.5, P1.5 ($n = 3$ brains/developmental stage), and P2.5 ($n = 1$) *En1^{Cre}; Rosa26^{Isl-GFP}* brains were quickly dissected on ice and immersed in 4% PFA for 12 hr before switching to PBS. 300- μm -thick coronal sections were cut on a vibratome (Leica VT 1200S). To increase the imaging depth, the sections were cleared following the ScaleSQ protocol (Hama et al., 2015). ScaleS4 buffer was used as a mounting medium (Hama et al., 2015), and spacers were placed on the slides to prevent compressing the sections. Nikon A1R inverted confocal was used to acquire z-stacks that

covered the entire extent of the thalamus for each brain, with a 20× objective (NA 0.75 Plan Apo VC). The achieved imaging depth in z ranged from 200 to 250 µm. The stacks were imported into Neurolucida software (MBF Bioscience) to trace the migratory morphology of GFP⁺ cells in the LGd, LP, and VP. On average, two sections covered the extent of these nuclei in the rostro-caudal dimension and the first time point when GFP⁺ cells were observed there was at E17.5. GFP⁺ cells were not traced in the PO and MG due to their low numbers in these nuclei in the juvenile and adult brains, and the ambiguity in delineating these regions anatomically in the embryonic brains. We did not observe GFP⁺ cells with neuronal morphology in any other TC regions (i.e. outside the FO and HO sensory thalamus) for all ages analysed. In the analysed regions (LGd, LP, and VP), all GFP⁺ somas were annotated using the semi-automated 'Soma' function. The leading processes were traced manually with the 'Tree' function, starting in the middle of the soma and until each process could be unequivocally identified or until the point of bifurcation, for all GFP⁺ cells with a clearly visible and identifiable leading process (44% of all GFP⁺ cells at E17.5, 30% at P0.5, 26% at P1.5, 14% at P2.5). The 3D coordinates for each leading process were then exported into Excel, and their orientation was expressed in the brain's coordinate system (x=L–M, y=V–M, z=C–R), as a vector joining the start and end point of the process, using a custom Python script and the Pandas (McKinney et al., 2010) and Numpy (van der Walt et al., 2011) libraries. Each vector was defined by its orientation in spherical coordinates (polar and azimuthal angle) and overall length. Population level orientation data for the LGd, LP, and VP at E17.5 and P0 was plotted as heat-maps, by binning cells according to their spherical coordinates. The bins were then integrated along each axis to reveal a dominant orientation (e.g. for the LGd, 66% and 69% of cells oriented dorso-ventrally and caudo-rostrally, respectively). Polar histograms of leading process orientation in the dorsal-ventral-lateral-medial plane were also produced.

Spatial clustering analysis

Unsupervised machine learning methods were used to investigate spatial organisation of Sox14⁺-Gad1⁺ and Sox14⁺Gad1⁺ cells. The 3D models of P14 Sox14^{GFP/+} thalamus generated with Neurolucida for NND analysis were again used to obtain the coordinates of all thalamic interneurons.

These data were analysed separately for each brain (n = 3) using a custom Python script, and partitioned into clusters using the k-means algorithm implemented in the library Scikit-Learn (Buitinck et al., 2013). The algorithm takes as input the expected number of clusters *k*.

Multiple values of *k* were tested, and evaluated using the silhouette coefficient metric of clustering performance (Rousseeuw, 1987), also implemented in Scikit-Learn. The silhouette coefficient is equal to the average ratio of distances between points within and between each cluster. More positive scores indicate coherent, well-separated clusters, whereas scores close to zero indicate overlapping clusters. The score was highest (0.472 ± 0.012) for *k* = 2, and the average fraction of all Sox14⁺ and Sox14⁺ cells in each of the resulting clusters was computed across all brains.

We also performed k-means clustering on the 3D distribution of Gad1⁺ cells obtained from ISH data from the Allen Mouse Brain Atlas. The silhouette score was again highest (0.512) for *k* = 2, and the resulting clusters have a spatial definition similar to those from the P14 Sox14^{GFP/+} thalamus.

Statistics

Comparison of distributions

The chi-squared test was used to test for significant differences in the thalamus-wide distribution of specific cell classes. This thalamus-wide comparison compensates for categorical errors arising from a degree of uncertainty in nuclear boundaries, as a result of variation in the sectioning plane and other factors.

For each distribution, average relative cell numbers were computed in Excel. A custom python script was used to compute the chi-squared statistic, and the corresponding p-value was computed using the chi-squared cumulative density function implemented in SciPy (Jones et al., 2001).

Change in interneuron numbers in the Sox14 knockout

This was tested for statistical significance using unpaired two-sample two-tailed t-test, comparing the Sox14 knockout to Sox14^{GFP/+} for each interneuron class separately (n = 3 brains/genotype).

Total interneuron numbers across all TC nuclei were compared and sampling was consistent between genotypes (each 10th thalamic section was analysed for each brain).

Identification of outliers

In the analysis of the Marmoset's thalamus, the fraction of GAD1 single positive cells is low in most TC nuclei tested, this low frequency is partly due to the lower efficiency of the SOX14 probe compared to the GAD1 and therefore a systematic error. In the LD and MD, however, the frequency of GAD1 single positive cells is higher. To demonstrate that values for these two TC nuclei are outliers, we applied the method described in *Motulsky and Brown, 2006*, implemented in GraphPad Prism software.

Acknowledgements

We thank the Wohl Cellular Imaging Centre, King's College London for support with imaging and image analysis software. We are grateful to Beatriz Rico, Monica Moissidis, and Patricia Hernandez at King's College London for the *Nkx2.1^{Cre}; Rosa26^{Isl-GFP}* and the *Lhx6^{Cre}; Rosa26^{Isl-GFP}* samples. We are grateful to the anonymous peer reviewers for their constructive and specific comments. This work was funded by the Biotechnology and Biological Sciences Research Council (BBSRC) grants BB/L020068/1 and BB/R007020/1 to AD and BB/R007659/1 to SB. Two-photon tomography was developed with Engineering and Physical Sciences Research Council (EPSRC) grant EP/J021199/1 to SRS.

Additional information

Funding

Funder	Grant reference number	Author
Biotechnology and Biological Sciences Research Council	BB/L020068/1	Alessio Delogu
Biotechnology and Biological Sciences Research Council	BB/R007020/1	Alessio Delogu
Biotechnology and Biological Sciences Research Council	BB/R007659/1	Stephen Brickley
Engineering and Physical Sciences Research Council	EP/J021199/1	Simon R Schultz
Engineering and Physical Sciences Research Council	EP/L016737/1	Gerald Moore

The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

Author contributions

Polona Jager, Conceptualization, Data curation, Software, Formal analysis, Investigation, Visualization, Methodology, Writing - original draft; Gerald Moore, Data curation, Software, Formal analysis, Methodology; Padraic Calpin, Data curation, Software, Methodology; Xhuljana Durmishi, Yoshiaki Kita, Yan Wang, Investigation, Methodology; Irene Salgarella, Data curation, Investigation; Lucy Menage, Dong Won Kim, Formal analysis, Investigation; Seth Blackshaw, Resources, Supervision; Simon R Schultz, Resources, Software, Supervision, Methodology; Stephen Brickley, Resources, Supervision, Funding acquisition; Tomomi Shimogori, Resources, Supervision, Methodology, Project administration; Alessio Delogu, Conceptualization, Data curation, Supervision, Funding acquisition, Investigation, Visualization, Methodology, Writing - original draft, Project administration, Writing - review and editing

Author ORCIDs

Polona Jager  <https://orcid.org/0000-0003-4484-9148>

Irene Salgarella  <https://orcid.org/0000-0003-3817-3321>

Simon R Schultz  <http://orcid.org/0000-0002-6794-5813>

Alessio Delogu  <https://orcid.org/0000-0002-4414-4714>

Ethics

Animal experimentation: Mice: Housing and experimental procedures were approved by the King's College London Ethical Committee and conformed to the regulations of the UK Home Office personal and project licences under the UK Animals (Scientific Procedures) 1986 Act. Marmoset: All experiments were conducted in accordance with the guidelines approved by the RIKEN Institutional Animal Care (W2020-2-022).

Decision letter and Author response

Decision letter <https://doi.org/10.7554/eLife.59272.sa1>

Author response <https://doi.org/10.7554/eLife.59272.sa2>

Additional files

Supplementary files

- Transparent reporting form

Data availability

All data generated or analysed during this study are included in the manuscript.

References

- Achim K, Peltopuro P, Lahti L, Tsai H-H, Zachariah A, Astrand M, Salminen M, Rowitch D, Partanen J. 2013. The role of Tal2 and Tal1 in the differentiation of midbrain GABAergic neuron precursors. *Biology Open* **2**:990–997. DOI: <https://doi.org/10.1242/bio.20135041>
- Achim K, Salminen M, Partanen J. 2014. Mechanisms regulating GABAergic neuron development. *Cellular and Molecular Life Sciences* **71**:1395–1415. DOI: <https://doi.org/10.1007/s00018-013-1501-3>, PMID: 24196748
- Albuixech-Crespo B, López-Blanch L, Burguera D, Maeso I, Sánchez-Arrones L, Moreno-Bravo JA, Somorjai I, Pascual-Anaya J, Puelles E, Bovolenta P, García-Fernández J, Puelles L, Irimia M, Ferran JL. 2017. Molecular regionalization of the developing amphioxus neural tube challenges major partitions of the vertebrate brain. *PLOS Biology* **15**:e2001573. DOI: <https://doi.org/10.1371/journal.pbio.2001573>, PMID: 28422959
- Anderson SA, Eisenstat DD, Shi L, Rubenstein JL. 1997a. Interneuron migration from basal forebrain to neocortex: dependence on dlx genes. *Science* **278**:474–476. DOI: <https://doi.org/10.1126/science.278.5337.474>, PMID: 9334308
- Anderson SA, Qiu M, Bulfone A, Eisenstat DD, Meneses J, Pedersen R, Rubenstein JL. 1997b. Mutations of the homeobox genes *Dlx-1* and *Dlx-2* disrupt the striatal subventricular zone and differentiation of late born striatal neurons. *Neuron* **19**:27–37. DOI: [https://doi.org/10.1016/S0896-6273\(00\)80345-1](https://doi.org/10.1016/S0896-6273(00)80345-1), PMID: 9247261
- Arcelli P, Frassoni C, Regondi MC, Biasi SD, Spreafico R. 1997. GABAergic neurons in mammalian thalamus: a marker of thalamic complexity? *Brain Research Bulletin* **42**:27–37. DOI: [https://doi.org/10.1016/S0361-9230\(96\)00107-4](https://doi.org/10.1016/S0361-9230(96)00107-4)
- Arendt D, Bertucci PY, Achim K, Musser JM. 2019. Evolution of neuronal types and families. *Current Opinion in Neurobiology* **56**:144–152. DOI: <https://doi.org/10.1016/j.conb.2019.01.022>
- Armstrong E. 1979. A quantitative comparison of the hominoid thalamus I specific sensory relay nuclei. *American Journal of Physical Anthropology* **51**:365–381. DOI: <https://doi.org/10.1002/ajpa.1330510308>
- Bakken TE, van Velthoven CTJ, Menon V, Hodge RD, Yao Z, Nguyen TN, Graybuck LT, Horwitz GD, Bertagnolli D, Goldy J. 2020. Single-cell RNA-seq uncovers shared and distinct axes of variation in dorsal LGN neurons in mice, non-human primates and humans. *bioRxiv*. DOI: <https://doi.org/10.1101/2020.11.05.367482>
- Baldwin MKL, Balaram P, Kaas JH. 2017. The evolution and functions of nuclei of the visual pulvinar in primates. *Journal of Comparative Neurology* **525**:3207–3226. DOI: <https://doi.org/10.1002/cne.24272>
- Bickford ME, Wei H, Eisenback MA, Chomsung RD, Slusarczyk AS, Dankowski AB. 2008. Synaptic organization of thalamocortical axon collaterals in the perigeniculate nucleus and dorsal lateral geniculate nucleus. *The Journal of Comparative Neurology* **508**:264–285. DOI: <https://doi.org/10.1002/cne.21671>
- Bickford ME, Slusarczyk A, Dilger EK, Krahe TE, Kucuk C, Guido W. 2010. Synaptic development of the mouse dorsal lateral geniculate nucleus. *Journal of Comparative Neurology* **518**:622–635. DOI: <https://doi.org/10.1002/cne.22223>
- Bolkan SS, Stujenske JM, Parnaudeau S, Spellman TJ, Rauffenbart C, Abbas AI, Harris AZ, Gordon JA, Kellendonk C. 2017. Thalamic projections sustain prefrontal activity during working memory maintenance. *Nature Neuroscience* **20**:987–996. DOI: <https://doi.org/10.1038/nn.4568>

- Braak H, Bachmann A. 1985. The percentage of projection neurons and interneurons in the human lateral geniculate nucleus. *Human Neurobiology* **4**:91–95. PMID: 4030427
- Buitinck L, Louppe G, Blondel M, Pedregosa F, Mueller A, Grisel O, Niculae V, Prettenhofer P, Gramfort A, Grobler J. 2013. API design for machine learning software: experiences from the scikit-learn project ECML PKDD Workshop: Languages for Data Mining and Machine Learning. arXiv. <https://arxiv.org/abs/1309.0238>.
- Bulfone A, Puelles L, Porteus MH, Frohman MA, Martin GR, Rubenstein JL. 1993. Spatially restricted expression of Dlx-1, Dlx-2 (Tes-1), Gbx-2, and Wnt-3 in the embryonic day 12.5 mouse forebrain defines potential transverse and longitudinal segmental boundaries. *The Journal of Neuroscience* **13**:3155–3172. DOI: <https://doi.org/10.1523/JNEUROSCI.13-07-03155.1993>, PMID: 7687285
- Butler AB. 2008. Evolution of the thalamus: a morphological and functional review. *Thalamus & Related Systems* **4**:35–58. DOI: <https://doi.org/10.1017/S1472928808000356>
- Cheong SK, Tailby C, Solomon SG, Martin PR. 2013. Cortical-Like receptive fields in the lateral geniculate nucleus of marmoset monkeys. *Journal of Neuroscience* **33**:6864–6876. DOI: <https://doi.org/10.1523/JNEUROSCI.5208-12.2013>
- Clascá F, Rubio-Garrido P, Jabaudon D. 2012. Unveiling the diversity of thalamocortical neuron subtypes. In *European Journal of Neuroscience* **35**:1524–1532. DOI: <https://doi.org/10.1111/j.1460-9568.2012.08033.x>
- Clemente-Perez A, Makinson SR, Higashikubo B, Brovarney S, Cho FS, Urry A, Holden SS, Wimer M, Dávid C, Fenno LE, Acsády L, Deisseroth K, Paz JT. 2017. Distinct thalamic reticular cell types differentially modulate normal and pathological cortical rhythms. *Cell Reports* **19**:2130–2142. DOI: <https://doi.org/10.1016/j.celrep.2017.05.044>
- Crone SA, Quinlan KA, Zagoraiou L, Droho S, Restrepo CE, Lundfald L, Endo T, Setlak J, Jessell TM, Kiehn O, Sharma K. 2008. Genetic ablation of V2a ipsilateral interneurons disrupts Left-Right locomotor coordination in mammalian spinal cord. *Neuron* **60**:70–83. DOI: <https://doi.org/10.1016/j.neuron.2008.08.009>
- D Blice M, Stocker C, Holzinger A. 2017. Augmentor: an image augmentation library for machine learning. *The Journal of Open Source Software* **2**:432. DOI: <https://doi.org/10.21105/joss.00432>
- Delogu A, Sellers K, Zagoraiou L, Bocianowska-Zbrog A, Mandal S, Guimera J, Rubenstein JLR, Sugden D, Jessell T, Lumsden A. 2012. Subcortical visual shell nuclei targeted by ipRGCs develop from a Sox14+ GABAergic progenitor and require Sox14 to regulate daily activity rhythms. *Neuron* **75**:648–662. DOI: <https://doi.org/10.1016/j.neuron.2012.06.013>
- Díaz C, Morales-Delgado N, Puelles L. 2014. Ontogenesis of peptidergic neurons within the genoarchitectonic map of the mouse hypothalamus. *Frontiers in Neuroanatomy* **8**:162. DOI: <https://doi.org/10.3389/fnana.2014.00162>, PMID: 25628541
- Evangelio M, García-Amado M, Clascá F. 2018. Thalamocortical projection neuron and interneuron numbers in the visual thalamic nuclei of the adult C57BL/6 mouse. *Frontiers in Neuroanatomy* **12**:27. DOI: <https://doi.org/10.3389/fnana.2018.00027>
- Fogarty M, Grist M, Gelman D, Marín O, Pachnis V, Kessaris N. 2007. Spatial genetic patterning of the embryonic neuroepithelium generates GABAergic interneuron diversity in the adult cortex. *Journal of Neuroscience* **27**:10935–10946. DOI: <https://doi.org/10.1523/JNEUROSCI.1629-07.2007>, PMID: 17928435
- Gelman DM, Martini FJ, Nóbrega-Pereira S, Pierani A, Kessaris N, Marín O. 2009. The embryonic preoptic area is a novel source of cortical GABAergic interneurons. *Journal of Neuroscience* **29**:9380–9389. DOI: <https://doi.org/10.1523/JNEUROSCI.0604-09.2009>, PMID: 19625528
- Gelman D, Griveau A, Dehorter N, Teissier A, Varela C, Pla R, Pierani A, Marín O. 2011. A wide diversity of cortical GABAergic interneurons derives from the embryonic preoptic area. *Journal of Neuroscience* **31**:16570–16580. DOI: <https://doi.org/10.1523/JNEUROSCI.4068-11.2011>, PMID: 22090484
- Golden JA, Cepko CL. 1996. Clones in the chick diencephalon contain multiple cell types and siblings are widely dispersed. *Development* **122**:65–78. PMID: 8565854
- Golding B, Pouchelon G, Bellone C, Murthy S, Di Nardo AA, Govindan S, Ogawa M, Shimogori T, Lüscher C, Dayer A, Jabaudon D. 2014. Retinal input directs the recruitment of inhibitory interneurons into thalamic visual circuits. *Neuron* **81**:1057–1069. DOI: <https://doi.org/10.1016/j.neuron.2014.01.032>
- Granda RH, Crossland WJ. 1989. GABA-like immunoreactivity of neurons in the chicken diencephalon and mesencephalon. *The Journal of Comparative Neurology* **287**:455–469. DOI: <https://doi.org/10.1002/cne.902870405>
- Groh A, Bokor H, Mease RA, Plattner VM, Hangya B, Stroh A, Deschenes M, Acsády L. 2014. Convergence of cortical and sensory driver inputs on single thalamocortical cells. *Cerebral Cortex* **24**:3167–3179. DOI: <https://doi.org/10.1093/cercor/bht173>
- Guillery RW. 1995. Anatomical evidence concerning the role of the thalamus in corticocortical communication: a brief review. *Journal of Anatomy* **187**:583–592. PMID: 8586557
- Guo ZV, Inagaki HK, Daie K, Druckmann S, Gerfen CR, Svoboda K. 2017. Maintenance of persistent activity in a frontal thalamocortical loop. *Nature* **545**:181–186. DOI: <https://doi.org/10.1038/nature22324>, PMID: 28467817
- Guo Q, Li JYH. 2019. Defining developmental diversification of diencephalon neurons through single cell gene expression profiling. *Development* **146**:dev174284. DOI: <https://doi.org/10.1242/dev.174284>, PMID: 30872278
- Halassa MM, Kastner S. 2017. Thalamic functions in distributed cognitive control. *Nature Neuroscience* **20**:1669–1679. DOI: <https://doi.org/10.1038/s41593-017-0020-1>
- Halley AC, Krubitzer L. 2019. Not all cortical expansions are the same: the coevolution of the neocortex and the dorsal thalamus in mammals. *Current Opinion in Neurobiology* **56**:78–86. DOI: <https://doi.org/10.1016/j.conb.2018.12.003>

- Hama H, Hioki H, Namiki K, Hoshida T, Kurokawa H, Ishidate F, Kaneko T, Akagi T, Saito T, Saido T, Miyawaki A. 2015. ScaleS: an optical clearing palette for biological imaging. *Nature Neuroscience* **18**:1518–1529. DOI: <https://doi.org/10.1038/nn.4107>
- Hayes SG, Murray KD, Jones EG. 2003. Two epochs in the development of γ -aminobutyric acidergic neurons in the ferret thalamus. *The Journal of Comparative Neurology* **463**:45–65. DOI: <https://doi.org/10.1002/cne.10749>
- Herkenham M. 1980. Laminar organization of thalamic projections to the rat neocortex. *Science* **207**:532–535. DOI: <https://doi.org/10.1126/science.7352263>
- Hippenmeyer S, Vrieseling E, Sigrist M, Portmann T, Laengle C, Ladle DR, Arber S. 2005. A developmental switch in the response of DRG neurons to ETS transcription factor signaling. *PLOS Biology* **3**:e159–. DOI: <https://doi.org/10.1371/journal.pbio.0030159>
- Hirsch JA, Wang X, Sommer FT, Martinez LM. 2015. How inhibitory circuits in the thalamus serve vision. *Annual Review of Neuroscience* **38**:309–329. DOI: <https://doi.org/10.1146/annurev-neuro-071013-014229>, PMID: 26154979
- Hubel DH, Wiesel TN. 1962. Receptive fields, binocular interaction and functional architecture in the cat's visual cortex. *The Journal of Physiology* **160**:106–154. DOI: <https://doi.org/10.1113/jphysiol.1962.sp006837>
- Huisman C, Cho H, Brock O, Lim SJ, Youn SM, Park Y, Kim S, Lee SK, Delogu A, Lee JW. 2019. Single cell transcriptome analysis of developing arcuate nucleus uncovers their key developmental regulators. *Nature Communications* **10**:3696. DOI: <https://doi.org/10.1038/s41467-019-11667-y>, PMID: 31420539
- Humphreys BD, Valerius MT, Kobayashi A, Mugford JW, Soeung S, Duffield JS, McMahon AP, Bonventre JV. 2008. Intrinsic epithelial cells repair the kidney after injury. *Cell Stem Cell* **2**:284–291. DOI: <https://doi.org/10.1016/j.stem.2008.01.014>, PMID: 18371453
- Hunnichutt BJ, Long BR, Kusefoglu D, Gertz KJ, Zhong H, Mao T. 2014. A comprehensive thalamocortical projection map at the mesoscopic level. *Nature Neuroscience* **17**:1276–1285. DOI: <https://doi.org/10.1038/nn.3780>
- Jager P, Ye Z, Yu X, Zagoraoui L, Prekop HT, Partanen J, Jessell TM, Wisden W, Brickley SG, Delogu A. 2016. Tectal-derived interneurons contribute to phasic and tonic inhibition in the visual thalamus. *Nature Communications* **7**:13579. DOI: <https://doi.org/10.1038/ncomms13579>, PMID: 27929058
- Jones E, Oliphant T, Peterson P. 2001. *SciPy: Open Source Scientific Tools for Python*.
- Jones E. 2002. Dichronous appearance and unusual origins of GABA neurons during development of the mammalian thalamus. *Thalamus and Related Systems* **1**:283–288. DOI: <https://doi.org/10.1017/S147292880200002X>
- Jones EG. 2007. *The Thalamus*. Cambridge University Press.
- Kenigfest NB, Repérant J, Rio J-P, Belekova MG, Tumanova NL, Ward R, Vesselkin NP, Herbin M, Chkeidze DD, Ozirskaya EV. 1995. Fine structure of the dorsal lateral geniculate nucleus of the turtle, *Emys orbicularis*: A Golgi, combined hrp tracing and GABA immunocytochemical study. *Journal of Comparative Neurology* **356**:595–614. DOI: <https://doi.org/10.1002/cne.903560409>
- Kenigfest NB, Repérant J, Rio J-P, Belekova MG, Ward R, Vesselkin NP, Miceli D, Herbin M. 1998. Retinal and cortical afferents to the dorsal lateral geniculate nucleus of the turtle, *Emys orbicularis*: a combined axonal tracing, glutamate, and GABA immunocytochemical electron microscopic study. In *The Journal of Comparative Neurology* **391**:470–490. DOI: [https://doi.org/10.1002/\(SICI\)1096-9861\(19980222\)391:4<470::AID-CNE5>3.0.CO;2-Y](https://doi.org/10.1002/(SICI)1096-9861(19980222)391:4<470::AID-CNE5>3.0.CO;2-Y)
- Kim DW, Liu K, Wang ZQ, Zhang YS, Bathini A, Brown MP, Lin SH, Washington PW, Sun C, Lindtner S. 2020. Gene regulatory networks controlling differentiation, survival, and diversification of hypothalamic Lhx6-expressing GABAergic neurons. *bioRxiv*. DOI: <https://doi.org/10.1101/2020.05.21.106963>
- Kimmel RA, Turnbull DH, Blanquet V, Wurst W, Loomis CA, Joyner AL. 2000. Two lineage boundaries coordinate vertebrate apical ectodermal ridge formation. *Genes & Development* **14**:1377–1389. PMID: 10837030
- Klein S, Staring M, Murphy K, Viergever MA, Pluim JP. 2010. Elastix: a toolbox for intensity-based medical image registration. *IEEE Transactions on Medical Imaging* **29**:196–205. DOI: <https://doi.org/10.1109/TMI.2009.2035616>, PMID: 19923044
- Lavdas AA, Grigoriou M, Pachnis V, Parnavelas JG. 1999. The medial ganglionic eminence gives rise to a population of early neurons in the developing cerebral cortex. *The Journal of Neuroscience* **19**:7881–7888. DOI: <https://doi.org/10.1523/JNEUROSCI.19-18-07881.1999>, PMID: 10479690
- Le TN, Zhou QP, Cobos I, Zhang S, Zagazewski J, Japoni S, Vriend J, Parkinson T, Du G, Rubenstein JL, Eisenstat DD. 2017. GABAergic interneuron differentiation in the basal forebrain is mediated through direct regulation of glutamic acid decarboxylase isoforms by dlx homeobox transcription factors. *The Journal of Neuroscience* **37**:8816–8829. DOI: <https://doi.org/10.1523/JNEUROSCI.2125-16.2017>, PMID: 28821666
- Lein ES, Hawrylycz MJ, Ao N, Ayres M, Bensinger A, Bernard A, Boe AF, Boguski MS, Brockway KS, Byrnes EJ, Chen L, Chen L, Chen TM, Chin MC, Chong J, Crook BE, Czaplinska A, Dang CN, Datta S, Dee NR, et al. 2007. Genome-wide atlas of gene expression in the adult mouse brain. *Nature* **445**:168–176. DOI: <https://doi.org/10.1038/nature05453>, PMID: 17151600
- Letinić K, Kostović I. 1997. Transient fetal structure, the gangliothalamic body, connects telencephalic germinal zone with all thalamic regions in the developing human brain. *The Journal of Comparative Neurology* **384**:373–395. DOI: [https://doi.org/10.1002/\(SICI\)1096-9861\(19970804\)384:3<373::AID-CNE5>3.0.CO;2-0](https://doi.org/10.1002/(SICI)1096-9861(19970804)384:3<373::AID-CNE5>3.0.CO;2-0), PMID: 9254034
- Letinić K, Rakic P. 2001. Telencephalic origin of human thalamic GABAergic neurons. *Nature Neuroscience* **4**:931–936. DOI: <https://doi.org/10.1038/nn0901-931>

- Ling S, Pratte MS, Tong F. 2015. Attention alters orientation processing in the human lateral geniculate nucleus. *Nature Neuroscience* **18**:496–498. DOI: <https://doi.org/10.1038/nn.3967>
- Liu JK, Ghattas I, Liu S, Chen S, Rubenstein JL. 1997. Dlx genes encode DNA-binding proteins that are expressed in an overlapping and sequential pattern during basal ganglia differentiation. *Developmental Dynamics* **210**:498–512. DOI: [https://doi.org/10.1002/\(SICI\)1097-0177\(199712\)210:4<498::AID-AJA12>3.0.CO;2-3](https://doi.org/10.1002/(SICI)1097-0177(199712)210:4<498::AID-AJA12>3.0.CO;2-3), PMID: 9415433
- Long JE, Garel S, Alvarez-Dolado M, Yoshikawa K, Osumi N, Alvarez-Buylla A, Rubenstein JL. 2007. Dlx-dependent and -independent regulation of olfactory bulb interneuron differentiation. *Journal of Neuroscience* **27**:3230–3243. DOI: <https://doi.org/10.1523/JNEUROSCI.5265-06.2007>, PMID: 17376983
- Long JE, Cobos I, Potter GB, Rubenstein JL. 2009a. Dlx1&2 and Mash1 transcription factors control MGE and CGE patterning and differentiation through parallel and overlapping pathways. *Cerebral Cortex* **19**:i96–i106. DOI: <https://doi.org/10.1093/cercor/bhp045>, PMID: 19386638
- Long JE, Swan C, Liang WS, Cobos I, Potter GB, Rubenstein JL. 2009b. Dlx1&2 and Mash1 transcription factors control striatal patterning and differentiation through parallel and overlapping pathways. *The Journal of Comparative Neurology* **512**:556–572. DOI: <https://doi.org/10.1002/cne.21854>, PMID: 19030180
- Marín O, Rubenstein JLR. 2001. A long, remarkable journey: tangential migration in the telencephalon. *Nature Reviews Neuroscience* **2**:780–790. DOI: <https://doi.org/10.1038/35097509>
- Martinez S, Alvarado-Mallart RM. 1989. Transplanted mesencephalic quail cells colonize selectively all primary visual nuclei of chick diencephalon: a study using heterotopic transplants. *Developmental Brain Research* **47**:263–274. DOI: [https://doi.org/10.1016/0165-3806\(89\)90181-8](https://doi.org/10.1016/0165-3806(89)90181-8)
- McKinney W, van der Walt S, Millman J. 2010. Data structures for statistical computing in Python. Proceedings of the 9th Python in Science Conference 51–56.
- Metin C, Alvarez C, Moudoux D, Vitalis T, Pieau C, Molnar Z. 2007. Conserved pattern of tangential neuronal migration during forebrain development. *Development* **134**:2815–2827. DOI: <https://doi.org/10.1242/dev.02869>
- Miller JA, Ding SL, Sunkin SM, Smith KA, Ng L, Szafer A, Ebbert A, Riley ZL, Royall JJ, Aiona K, Arnold JM, Bennet C, Bertagnolli D, Brouner K, Butler S, Caldejon S, Carey A, Cuhacian C, Dalley RA, Dee N, et al. 2014. Transcriptional landscape of the prenatal human brain. *Nature* **508**:199–206. DOI: <https://doi.org/10.1038/nature13185>, PMID: 24695229
- Mo A, Mukamel EA, Davis FP, Luo C, Henry GL, Picard S, Urich MA, Nery JR, Sejnowski TJ, Lister R, Eddy SR, Ecker JR, Nathans J. 2015. Epigenomic signatures of neuronal diversity in the mammalian brain. *Neuron* **86**:1369–1384. DOI: <https://doi.org/10.1016/j.neuron.2015.05.018>
- Monory K, Massa F, Egertová M, Eder M, Blaudzun H, Westenbroek R, Kelsch W, Jacob W, Marsch R, Ekker M, Long J, Rubenstein JL, Goebbels S, Nave K-A, During M, Klugmann M, Wölfel B, Dodt H-U, Zieglgänsberger W, Wotjak CT, et al. 2006. The endocannabinoid system controls key epileptogenic circuits in the hippocampus. *Neuron* **51**:455–466. DOI: <https://doi.org/10.1016/j.neuron.2006.07.006>
- Montero VM. 1987. Ultrastructural identification of synaptic terminals from the axon of type 3 interneurons in the cat lateral geniculate nucleus. *The Journal of Comparative Neurology* **264**:268–283. DOI: <https://doi.org/10.1002/cne.902640210>
- Morales-Delgado N, Castro-Robles B, Ferrán JL, Martínez-de-la-Torre M, Puellas L, Díaz C. 2014. Regionalized differentiation of CRH, TRH, and GHRH peptidergic neurons in the mouse hypothalamus. *Brain Structure and Function* **219**:1083–1111. DOI: <https://doi.org/10.1007/s00429-013-0554-2>, PMID: 24337236
- Motulsky HJ, Brown RE. 2006. Detecting outliers when fitting data with nonlinear regression - a new method based on robust nonlinear regression and the false discovery rate. *BMC Bioinformatics* **7**:123. DOI: <https://doi.org/10.1186/1471-2105-7-123>, PMID: 16526949
- Newman EA, Kim DW, Wan J, Wang J, Qian J, Blackshaw S. 2018. Foxd1 is required for terminal differentiation of anterior hypothalamic neuronal subtypes. *Developmental Biology* **439**:102–111. DOI: <https://doi.org/10.1016/j.ydbio.2018.04.012>, PMID: 29679559
- Paredes MF, James D, Gil-Perotin S, Kim H, Cotter JA, Ng C, Sandoval K, Rowitch DH, Xu D, McQuillen PS, Garcia-Verdugo J-M, Huang EJ, Alvarez-Buylla A. 2016. Extensive migration of young neurons into the infant human frontal lobe. *Science* **354**:aaf7073. DOI: <https://doi.org/10.1126/science.aaf7073>
- Pasik P, Pasik T. 1976. Synapses between interneurons in the lateral geniculate nucleus of monkeys. *Experimental Brain Research* **25**:1–13. DOI: <https://doi.org/10.1007/BF00237322>
- Penny GR, Conley M, Schmechel DE, Diamond IT. 1984. The distribution of glutamic acid decarboxylase immunoreactivity in the diencephalon of the opossum and rabbit. *The Journal of Comparative Neurology* **228**:38–56. DOI: <https://doi.org/10.1002/cne.902280106>
- Piscopo DM, El-Danaf RN, Huberman AD, Niell CM. 2013. Diverse visual features encoded in mouse lateral geniculate nucleus. *Journal of Neuroscience* **33**:4642–4656. DOI: <https://doi.org/10.1523/JNEUROSCI.5187-12.2013>
- Pleasure SJ, Anderson S, Hevner R, Bagri A, Marín O, Lowenstein DH, Rubenstein JL. 2000. Cell migration from the ganglionic eminences is required for the development of hippocampal GABAergic interneurons. *Neuron* **28**:727–740. DOI: [https://doi.org/10.1016/S0896-6273\(00\)00149-5](https://doi.org/10.1016/S0896-6273(00)00149-5), PMID: 11163262
- Prekop HT, Kroiss A, Rook V, Zagoraoui L, Jessell TM, Fernandes C, Delogu A, Wingate RJT. 2018. Sox14 is required for a specific subset of Cerebello-Olivary projections. *The Journal of Neuroscience* **38**:9539–9550. DOI: <https://doi.org/10.1523/JNEUROSCI.1456-18.2018>, PMID: 30242051

- Pritz MB, Stritzel ME. 1994. Morphological and GAD immunocytochemical properties of the dorsal lateral geniculate nucleus in a reptile. In *Brain Research Bulletin* **33**:723–726. DOI: [https://doi.org/10.1016/0361-9230\(94\)90239-9](https://doi.org/10.1016/0361-9230(94)90239-9), PMID: 8193926
- Puelles L, Rubenstein JLR. 2003. Forebrain gene expression domains and the evolving prosomeric model. *Trends in Neurosciences* **26**:469–476. DOI: [https://doi.org/10.1016/S0166-2236\(03\)00234-0](https://doi.org/10.1016/S0166-2236(03)00234-0)
- Ragan T, Kadiri LR, Venkataraju KU, Bahlmann K, Sutin J, Taranda J, Arganda-Carreras I, Kim Y, Seung HS, Osten P. 2012. Serial two-photon tomography for automated ex vivo mouse brain imaging. *Nature Methods* **9**:255–258. DOI: <https://doi.org/10.1038/nmeth.1854>, PMID: 22245809
- Rakić P, Sidman RL. 1969. Telencephalic origin of pulvinar neurons in the fetal human brain. *Zeitschrift Für Anatomie Und Entwicklungsgeschichte* **129**:53–82. DOI: <https://doi.org/10.1007/BF00521955>
- Rausell E, Jones EG. 1991. Histochemical and immunocytochemical compartments of the thalamic VPM nucleus in monkeys and their relationship to the representational map. *The Journal of Neuroscience* **11**:210–225. DOI: <https://doi.org/10.1523/JNEUROSCI.11-01-00210.1991>, PMID: 1846010
- Rikhye RV, Gilra A, Halassa MM. 2018a. Thalamic regulation of switching between cortical representations enables cognitive flexibility. *Nature Neuroscience* **21**:1753–1763. DOI: <https://doi.org/10.1038/s41593-018-0269-z>
- Rikhye RV, Wimmer RD, Halassa MM. 2018b. Toward an integrative theory of thalamic function. *Annual Review of Neuroscience* **41**:163–183. DOI: <https://doi.org/10.1146/annurev-neuro-080317-062144>
- Rio J-P, Reperant J, Ward R, Miceli D, Medina M. 1992. Evidence of GABA-immunopositive neurons in the dorsal part of the lateral geniculate nucleus of reptiles: morphological correlates with interneurons. *Neuroscience* **47**:395–407. DOI: [https://doi.org/10.1016/0306-4522\(92\)90254-Y](https://doi.org/10.1016/0306-4522(92)90254-Y)
- Ronneberger O, Brox T. 2015. *U-Net: Convolutional Networks for Biomedical Image Segmentation*. Springer.
- Rousseeuw PJ. 1987. Silhouettes: a graphical aid to the interpretation and validation of cluster analysis. *Journal of Computational and Applied Mathematics* **20**:53–65. DOI: [https://doi.org/10.1016/0377-0427\(87\)90125-7](https://doi.org/10.1016/0377-0427(87)90125-7)
- Rubio-Garrido P, Pérez-de-Manzo F, Porrero C, Galazo MJ, Clascá F. 2009. Thalamic input to distal apical dendrites in neocortical layer 1 is massive and highly convergent. *Cerebral Cortex* **19**:2380–2395. DOI: <https://doi.org/10.1093/cercor/bhn259>
- Saalmann YB, Kastner S. 2011. Cognitive and perceptual functions of the visual thalamus. *Neuron* **71**:209–223. DOI: <https://doi.org/10.1016/j.neuron.2011.06.027>
- Saunders A, Macosko EZ, Wysoker A, Goldman M, Krien FM, de Rivera H, Bien E, Baum M, Bortolin L, Wang S, Goeva A, Nemesh J, Kamitaki N, Brumbaugh S, Kulp D, McCarroll SA. 2018. Molecular diversity and specializations among the cells of the adult mouse brain. In *Cell* **174**:1015–1030. DOI: <https://doi.org/10.1016/j.cell.2018.07.028>, PMID: 30096299
- Schindelin J, Rueden CT, Hiner MC, Eliceiri KW. 2015. The ImageJ ecosystem: an open platform for biomedical image analysis. *Molecular Reproduction and Development* **82**:518–529. DOI: <https://doi.org/10.1002/mrd.22489>, PMID: 26153368
- Schmitt LI, Wimmer RD, Nakajima M, Happ M, Mofakham S, Halassa MM. 2017. Thalamic amplification of cortical connectivity sustains attentional control. *Nature* **545**:219–223. DOI: <https://doi.org/10.1038/nature22073>
- Seabrook TA, El-Danaf RN, Krahe TE, Fox MA, Guido W. 2013a. Retinal input regulates the timing of corticogeniculate innervation. *Journal of Neuroscience* **33**:10085–10097. DOI: <https://doi.org/10.1523/JNEUROSCI.5271-12.2013>, PMID: 23761904
- Seabrook TA, Krahe TE, Govindaiah G, Guido W. 2013b. Interneurons in the mouse visual thalamus maintain a high degree of retinal convergence throughout postnatal development. *Neural Development* **8**:24. DOI: <https://doi.org/10.1186/1749-8104-8-24>
- Sellers K, Zyka V, Lumsden AG, Delogu A. 2014. Transcriptional control of GABAergic neuronal subtype identity in the thalamus. *Neural Development* **9**:14. DOI: <https://doi.org/10.1186/1749-8104-9-14>, PMID: 24929424
- Sgaier SK, Lao Z, Villanueva MP, Berenshteyn F, Stephen D, Turnbull RK, Joyner AL. 2007. Genetic subdivision of the tectum and cerebellum into functionally related regions based on differential sensitivity to engrailed proteins. *Development* **134**:2325–2335. DOI: <https://doi.org/10.1242/dev.000620>
- Shatz CJ. 1996. Emergence of order in visual system development. *PNAS* **93**:602–608. DOI: <https://doi.org/10.1073/pnas.93.2.602>, PMID: 8570602
- Sherman SM. 2004. Interneurons and triadic circuitry of the thalamus. *Trends in Neurosciences* **27**:670–675. DOI: <https://doi.org/10.1016/j.tins.2004.08.003>
- Sherman SM. 2016. Thalamus plays a central role in ongoing cortical functioning. *Nature Neuroscience* **19**:533–541. DOI: <https://doi.org/10.1038/nn.4269>
- Sherman SM, Guillery RW. 2002. The role of the thalamus in the flow of information to the cortex. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences* **357**:1695–1708. DOI: <https://doi.org/10.1098/rstb.2002.1161>
- Shi W, Xianyu A, Han Z, Tang X, Li Z, Zhong H, Mao T, Huang K, Shi S-H. 2017. Ontogenetic establishment of order-specific nuclear organization in the mammalian thalamus. *Nature Neuroscience* **20**:516–528. DOI: <https://doi.org/10.1038/nn.4519>
- Shimamura K, Hartigan DJ, Martinez S, Puelles L, Rubenstein JL. 1995. Longitudinal organization of the anterior neural plate and neural tube. *Development* **121**:3923–3933.
- Shimogori T, Lee DA, Miranda-Angulo A, Yang Y, Wang H, Jiang L, Yoshida AC, Kataoka A, Mashiko H, Avetisyan M, Qi L, Qian J, Blackshaw S. 2010. A genomic atlas of mouse hypothalamic development. *Nature Neuroscience* **13**:767–775. DOI: <https://doi.org/10.1038/nn.2545>

- Simeone A**, Acampora D, Pannese M, D'Esposito M, Stornaiuolo A, Gulisano M, Mallamaci A, Kastury K, Druck T, Huebner K. 1994. Cloning and characterization of two members of the vertebrate *Dlx* gene family. *PNAS* **91**: 2250–2254. DOI: <https://doi.org/10.1073/pnas.91.6.2250>
- Sousa VH**, Miyoshi G, Hjerling-Leffler J, Karayannis T, Fishell G. 2009. Characterization of *Nkx6-2*-Derived Neocortical Interneuron Lineages. *Cerebral Cortex* **19**:i1–i10. DOI: <https://doi.org/10.1093/cercor/bhp038>
- Stephan H**, Frahm H, Baron G. 1981. New and Revised Data on Volumes of Brain Structures in Insectivores and Primates. *Folia Primatologica* **35**:1–29. DOI: <https://doi.org/10.1159/000155963>
- Sugahara F**, Pascual-Anaya J, Oisi Y, Kuraku S, Aota S, Adachi N, Takagi W, Hirai T, Sato N, Murakami Y, Kuratani S. 2016. Evidence from cyclostomes for complex regionalization of the ancestral vertebrate brain. *Nature* **531**:97–100. DOI: <https://doi.org/10.1038/nature16518>
- Sugahara F**, Murakami Y, Pascual-Anaya J, Kuratani S. 2017. Reconstructing the ancestral vertebrate brain. *Development, Growth & Differentiation* **59**:163–174. DOI: <https://doi.org/10.1111/dgd.12347>
- Sussel L**, Marin O, Kimura S, Rubenstein JL. 1999. Loss of *Nkx2.1* homeobox gene function results in a ventral to dorsal molecular respecification within the basal telencephalon: evidence for a transformation of the pallidum into the striatum. *Development* **126**:3359–3370.
- Tasic B**, Menon V, Nguyen TN, Kim TK, Jarsky T, Yao Z, Levi B, Gray LT, Sorensen SA, Dolbeare T, Bertagnolli D, Goldy J, Shapovalova N, Parry S, Lee C, Smith K, Bernard A, Madisen L, Sunkin SM, Hawrylycz M, et al. 2016. Adult mouse cortical cell taxonomy revealed by single cell transcriptomics. *Nature Neuroscience* **19**:335–346. DOI: <https://doi.org/10.1038/nn.4216>
- Tasic B**, Yao Z, Graybuck LT, Smith KA, Nguyen TN, Bertagnolli D, Goldy J, Garren E, Economo MN, Viswanathan S, Penn O, Bakken T, Menon V, Miller J, Fong O, Hirokawa KE, Lathia K, Rimorin C, Tieu M, Larsen R, et al. 2018. Shared and distinct transcriptomic cell types across neocortical areas. *Nature* **563**:72–78. DOI: <https://doi.org/10.1038/s41586-018-0654-5>
- Thompson AD**, Picard N, Min L, Fagiolini M, Chen C. 2016. Cortical Feedback Regulates Feedforward Retinogeniculate Refinement. *Neuron* **91**:1021–1033. DOI: <https://doi.org/10.1016/j.neuron.2016.07.040>
- Tosches MA**, Yamawaki TM, Naumann RK, Jacobi AA, Tushev G, Laurent G. 2018. Evolution of pallium, Hippocampus, and cortical cell types revealed by single-cell transcriptomics in reptiles. *Science* **360**:881–888. DOI: <https://doi.org/10.1126/science.aar4237>, PMID: 29724907
- Tremblay R**, Lee S, Rudy B. 2016. GABAergic Interneurons in the Neocortex: From Cellular Properties to Circuits. *Neuron* **91**:260–292. DOI: <https://doi.org/10.1016/j.neuron.2016.06.033>
- Van der Loos H**, Woolsey TA. 1973. Somatosensory cortex: structural alterations following early injury to sense organs. *Science* **179**:395–398. DOI: <https://doi.org/10.1126/science.179.4071.395>, PMID: 4682966
- van der Walt S**, Colbert SC, Varoquaux G. 2011. The NumPy array: a structure for efficient numerical computation. *Computing in Science & Engineering* **13**:22–30. DOI: <https://doi.org/10.1109/MCSE.2011.37>
- Veenman CL**, Reiner A. 1994. The distribution of GABA-containing perikarya, fibers, and terminals in the forebrain and midbrain of pigeons, with particular reference to the basal ganglia and its projection targets. *The Journal of Comparative Neurology* **339**:209–250. DOI: <https://doi.org/10.1002/cne.903390205>
- Wang Y**, Dye CA, Sohal V, Long JE, Estrada RC, Roztocil T, Lufkin T, Deisseroth K, Baraban SC, Rubenstein JLR. 2010. *Dlx5* and *Dlx6* Regulate the Development of Parvalbumin-Expressing Cortical Interneurons. *Journal of Neuroscience* **30**:5334–5345. DOI: <https://doi.org/10.1523/JNEUROSCI.5963-09.2010>
- Watakabe A**, Ohsawa S, Hashikawa T, Yamamori T. 2006. Binding and complementary expression patterns of semaphorin 3E and plexin D1 in the mature neocortices of mice and monkeys. *The Journal of Comparative Neurology* **499**:258–273. DOI: <https://doi.org/10.1002/cne.21106>
- Wichterle H**, Turnbull DH, Nery S, Fishell G, Alvarez-Buylla A. 2001. In utero fate mapping reveals distinct migratory pathways and fates of neurons born in the mammalian basal forebrain. *Development* **128**:3759–3771. PMID: 11585802
- Wong SZH**, Scott EP, Mu W, Guo X, Borgenheimer E, Freeman M, Ming G-li, Wu Q-F, Song H, Nakagawa Y. 2018. In vivo clonal analysis reveals spatiotemporal regulation of thalamic nucleogenesis. *PLOS Biology* **16**: e2005211. DOI: <https://doi.org/10.1371/journal.pbio.2005211>
- Xu Q**, Cobos I, De La Cruz E, Rubenstein JL, Anderson SA. 2004. Origins of Cortical Interneuron Subtypes. *Journal of Neuroscience* **24**:2612–2622. DOI: <https://doi.org/10.1523/JNEUROSCI.5667-03.2004>
- Xu Q**, Tam M, Anderson SA. 2008. Fate mapping *Nkx2.1*-lineage cells in the mouse telencephalon. *The Journal of Comparative Neurology* **506**:16–29. DOI: <https://doi.org/10.1002/cne.21529>
- Yun K**, Fischman S, Johnson J, Hrabe de Angelis M, Weinmaster G, Rubenstein JL. 2002. Modulation of the notch signaling by *Mash1* and *Dlx1/2* regulates sequential specification and differentiation of progenitor cell types in the subcortical telencephalon. *Development* **129**:5029–5040. PMID: 12397111
- Zeater N**, Cheong SK, Solomon SG, Dreher B, Martin PR. 2015. Binocular visual responses in the primate lateral geniculate nucleus. *Current Biology* **25**:3190–3195. DOI: <https://doi.org/10.1016/j.cub.2015.10.033>, PMID: 26778654
- Zhao Y**, Flandin P, Long JE, Cuesta MD, Westphal H, Rubenstein JLR. 2008. Distinct molecular pathways for development of telencephalic interneuron subtypes revealed through analysis of *Lhx6* mutants. *The Journal of Comparative Neurology* **510**:79–99. DOI: <https://doi.org/10.1002/cne.21772>